



US 20220411746A1

(19) **United States**

(12) **Patent Application Publication**
MIRELES et al.

(10) **Pub. No.: US 2022/0411746 A1**

(43) **Pub. Date: Dec. 29, 2022**

(54) **PHAGE-RESISTANT MICROORGANISMS**

(86) PCT No.: **PCT/IB2019/001262**

(71) Applicant: **INNOVACION Y DESARROLLO DE ENERGIA ALFA SUSTENTABLE S.A. DE C.V.**, Nuevo Leon, Mexico (MX)

§ 371 (c)(1),
(2) Date: **May 18, 2022**

Publication Classification

(72) Inventors: **Ivan Alejandro de la Pena MIRELES**, Monterrey, Nuevo Leon (MX); **Claudio Garibay ORIJEL**, Metepec, Estado de Mexico (MX); **Liliana Dondiego RODRIGUEZ**, Toluca, Estado de Mexico (MX); **Javier Acedo ZUNIGA**, Ciudad de Mexico (MX)

(51) **Int. Cl.**
C12N 1/20 (2006.01)
C12Q 1/10 (2006.01)
C12N 15/70 (2006.01)

(52) **U.S. Cl.**
CPC **C12N 1/205** (2021.05); **C12Q 1/10** (2013.01); **C12N 15/70** (2013.01); **C12R 2001/19** (2021.05)

(73) Assignee: **INNOVACION Y DESARROLLO DE ENERGIA ALFA SUSTENTABLE S.A. DE C.V.**, Nuevo Leon, Mexico (MX)

(57) **ABSTRACT**

(21) Appl. No.: **17/777,878**

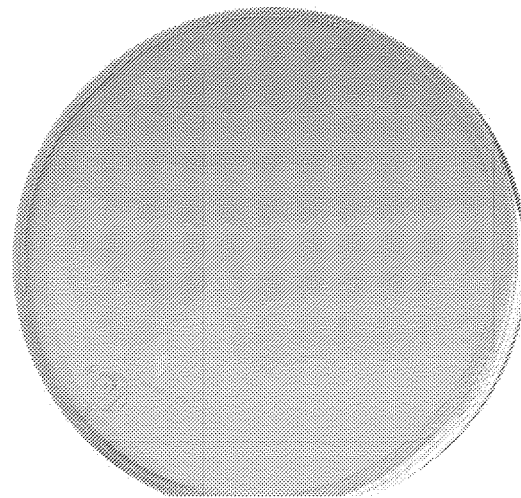
Genetically modified microorganisms which are resistant to infection by bacteriophages and that retain their kinetic parameters and methods of making the same.

(22) PCT Filed: **Nov. 27, 2019**

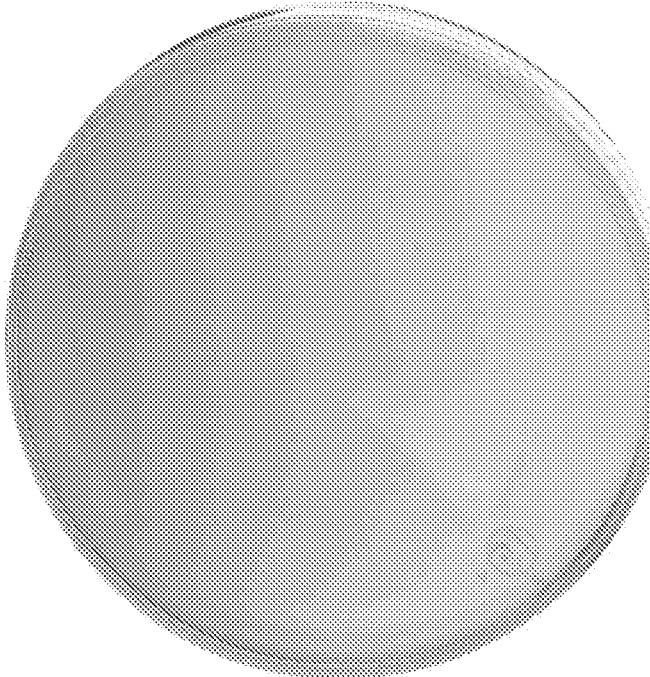
Specification includes a Sequence Listing.



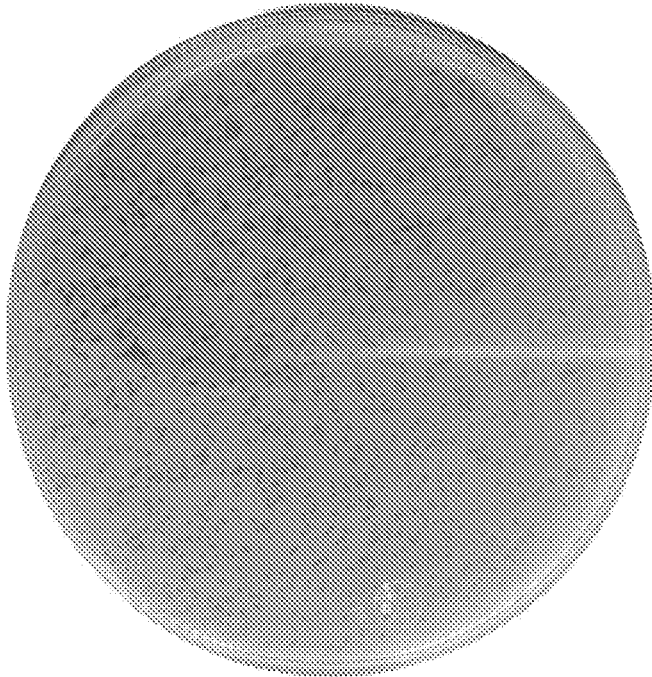
E. coli K-12 wild type



E. coli LCT-BF-01



E. coli LCT-BF-01



E. coli K-12 wild type

Figure 1.

Figure 2

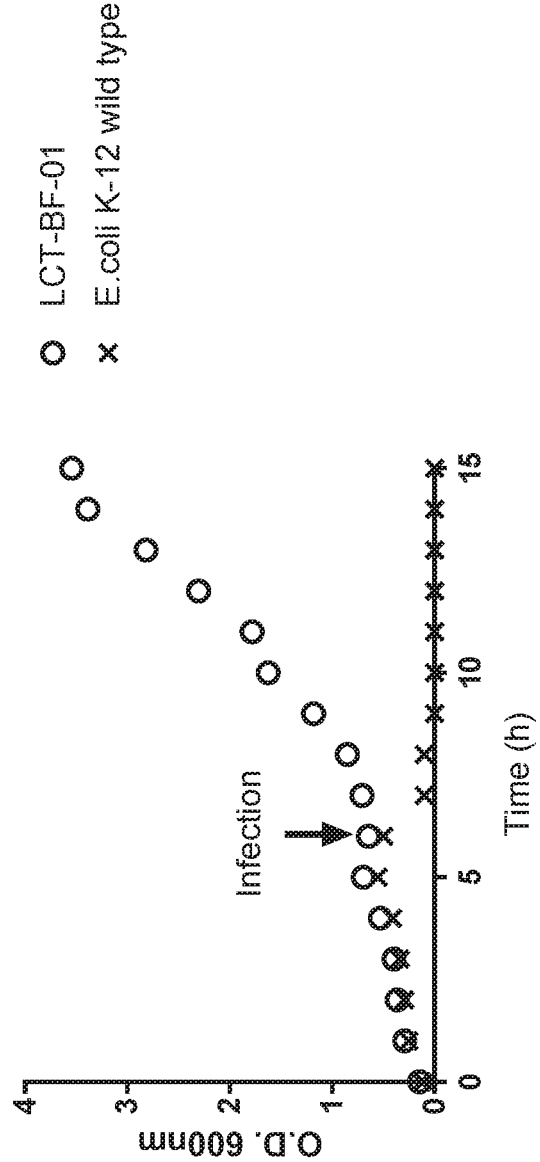


Figure. 3

Alignment tfaD (aminoacid change A -> T)

>TfaD Protein alignment Alignment of 2 sequences: TfaD wildtype, TfaD mutated

Score = 757.0, Identities = 145/146 (99%), Positives = 145/146 (99%), Gaps = 0/146 (0%)

```

TfaD wildtype      1 IPAGFVAVFNSDESSWHLVEDHRGKTVYDVASGDALFISELGPLPENVTWLSPEGEFQK
W 60
Consensus          IPAGFVAVFNSDESSWHLVEDHRGKTVYDVASGDALFISELGPLPENVTWLSPEGEFQK
W
TfaD mutated      1 IPAGFVAVFNSDESSWHLVEDHRGKTVYDVASGDALFISELGPLPENVTWLSPEGEFQK
W 60

TfaD wildtype     61 NGTAWVKD EAEKLFRIREAETKNSLMQVASEHIAPLQDAVDLEIATEEETSLLEAWK
K 120
Consensus         NGTAWVKD EAEKLFRIREAETKNSLMQVASEHIAPLQDAVDLEIATEEETSLLEAWK
K
TfaD mutated     61 NGTAWVKD EAEKLFRIREAETKNSLMQVASEHIAPLQDAVDLEIATEEETSLLEAWK
K 120

TfaD wildtype    121 YRVLLNRVDTSTAPDIEWPTNPVRE* 146
Consensus        YRVLLNRVDTSTAPDIEWPTNPVRE*
TfaD mutated    121 YRVLLNRVDTSTAPDIEWPTNPVRE* 146
    
```

Figure. 4

>TfaD Nucleotide alignment Alignment of 2 sequences: TfaD wildtype, TfaD mutated

Score = 2063.764, Identities = 429/438 (97%), Positives = 429/438 (97%), Gaps = 0/438 (0%)

```

TfaD wildtype      1 ATTCCAGCTGGCTTCGTGGCTGTTTTCAACAGTGATGAGTCATCGTGGCATCTCGTTGA
A 60
Consensus          ATTCCAGCTGGCTTCGTGGCTGTTTTCAACAGTGATGAGTCATCGTGGCATCTCGTTGA
A
TfaD mutated       1 ATTCCAGCTGGCTTCGTGGCTGTTTTCAACAGTGATGAGTCATCGTGGCATCTCGTTGA
A 60

TfaD wildtype     61 GATCATCGGGGTAAAACGGTTTATGACGTAGCGTCAGGGGACGCGTTATTTATTTCTGA
A 120
Consensus          GATCATCGGGGTAAAACGGTTTATGACGT GC TC GG GACGCGTTATTTATTTCTGA
A
TfaD mutated      61 GATCATCGGGGTAAAACGGTTTATGACGTGGCTTCCGGCGACGCGTTATTTATTTCTGA
A 120

TfaD wildtype     121 CTCGGTCCGTTACCGGAAAATGTTACCTGGTTATCGCCGGAAGGGGAGTTTCAGAAGTG
G 180
Consensus          CTCGGTCCGTTACCGGAAAATGTTACCTGGTTATCGCCGGAAGGGGAGTTTCAGAAGTG
G
TfaD mutated      121 CTCGGTCCGTTACCGGAAAATGTTACCTGGTTATCGCCGGAAGGGGAGTTTCAGAAGTG
G 180

TfaD wildtype     181 AACGGTACAGCCTGGGTGAAAGATGCGAGAAGCAGAAAAACTGTTCCGGATTCGGGAGGC
G 240
Consensus          AACGG ACAGCCTGGGTGAA GAT C GAAGCAGAAAAACTGTTCCGGAT CGGGAGGC
G
TfaD mutated      181 AACGGCACAGCCTGGGTGAAAGGATGCGGAAGCAGAAAAACTGTTCCGGATCCGGGAGGC
G 240

TfaD wildtype     241 GAAGAAACAAAAACAGCCTGATGCAGGTAGCCAGTGAGCATATTGCGCCACTTCAGGA
T 300
Consensus          GAAGAAACAAAAACAGCCTGATGCAGGTAGCCAGTGAGCATATTGCGCCACTTCAGGA
T
TfaD mutated      241 GAAGAAACAAAAACAGCCTGATGCAGGTAGCCAGTGAGCATATTGCGCCACTTCAGGA
T 300

TfaD wildtype     301 GCTGTAGATCTGGAAATCGCAACGGAGGAAGAAAACCTCATTGCTGGAAGCCTGGAAAAA
A 360
Consensus          GCTGTAGATCTGGAAATCGCAACGGAGGAAGAAAACCTCATTGCTGGAAGCCTGGAAAAA
A
TfaD mutated      301 GCTGTAGATCTGGAAATCGCAACGGAGGAAGAAAACCTCATTGCTGGAAGCCTGGAAAAA
A 360

TfaD wildtype     361 TATCGGGTGTGCTGAACCGTGTGATAACATCAACTGCACCTGATATTGAGTGGCCTAC
G 420
Consensus          TATCGGGTGTGCTGAACCGTGTGATAACATCAACTGCACCTGATATTGAGTGGCCTAC
G
TfaD mutated      361 TATCGGGTGTGCTGAACCGTGTGATAACATCAACTGCACCTGATATTGAGTGGCCTAC
G 420

TfaD wildtype     421 AACCTGTCAGGGAGTAA 438
Consensus          AACCTGTCAGGGAGTAA
TfaD mutated      421 AACCTGTCAGGGAGTAA 438
    
```

Figure. 5

Alignment yejO (aminoacid change W -> R)

>yejO Protein alignment Alignment of 2 sequences: yejO wildtype, yejO mutated

Score = 6358.0, Identities = 1262/1263 (99%), Positives = 1262/1263 (99%), Gaps = 0/1263 (0%)

yejO wildtype SSK 60 Consensus SSK	1	MHQSGSVSLCRSAISVLVATALK W KVRTSP*YEIMFVIWSHRTGFIMSHQLTFADSEF
yejO mutated SSK 60	1	MHQSGSVSLCRSAISVLVATALK W KVRTSP*YEIMFVIWSHRTGFIMSHQLTFADSEF
yejO wildtype SDG 120 Consensus SDG	61	RRQTRKEIFLSRMEQILPWQNMVEVI EPPFYPKAGNGRRPYPLETMLRIHCMQHWNL
yejO mutated SDG 120	61	RRQTRKEIFLSRMEQILPWQNMVEVI EPPFYPKAGNGRRPYPLETMLRIHCMQHWNL
yejO wildtype GVM 180 Consensus GVM	121	AMEDALYEIASMRLFARLSLDSALPDRTTIMNFRHLL W EQHQLARQLFKTINRWLAEA
yejO mutated GVM 180	121	AMEDALYEIASMRLFARLSLDSALPDRTTIMNFRHLL W EQHQLARQLFKTINRWLAEA
yejO wildtype LVT 240 Consensus LVT	181	MTQGTLVDATII EAPSSTKNKEQQRDPEMHQTKKGNQWHFGMKAHIGVDAKSGLTHS
yejO mutated LVT 240	181	MTQGTLVDATII EAPSSTKNKEQQRDPEMHQTKKGNQWHFGMKAHIGVDAKSGLTHS
yejO wildtype QHP 300 Consensus QHP	241	TAANEHDLNQLGNLLHGEEQFVSADAGYQGAPQREELAEVDVDWLIAERPGKVRTLK
yejO mutated QHP 300	241	TAANEHDLNQLGNLLHGEEQFVSADAGYQGAPQREELAEVDVDWLIAERPGKVRTLK
yejO wildtype FRA 360 Consensus FRA	301	RKNKTAINIEYMKASIRARVEHPFRI IKRQFGFVKARYKGLLKNDNQLAMLFTLANL
yejO mutated FRA 360	301	RKNKTAINIEYMKASIRARVEHPFRI IKRQFGFVKARYKGLLKNDNQLAMLFTLANL
yejO wildtype SHL 420 Consensus SHL	361	DQMIRQWERSH*KLGI TP*MAKKRSK*ADSRHLREKKS W AQT*RNEMTESAEKNFPAY
yejO mutated SHL 420	361	DQMIRQWERSH*KLGI TP*MAKKRSK*ADSRHLREKKS W AQT*RNEMTESAEKNFPAY
yejO wildtype LTE 480 Consensus LTE	421	PYIHP*HWHQLLSMVRQLMVL W SWKKISSWFMGPPI IRKSILAENSI *KNLV*VI I LK
yejO mutated LTE 480	421	PYIHP*HWHQLLSMVRQLMVL W SWKKISSWFMGPPI IRKSILAENSI *KNLV*VI I LK
yejO wildtype	481	GISTLK*MAPQNTQY*MTVIKLFK W VARQTRLRSIMVCYRFMAQRMIPRLKAGA*SL

KKM 540 Consensus KKM yej0 mutated KKM 540	481	GISTLK*MAPQNTQY*MTVIKLFKWVARQTRLRSIMVCYRFMAQRMI PRLKAGA*SL
yej0 wildtype LII 600 Consensus LII yej0 mutated LII 600	541	GGPSLSLSKREDYWRLKRGDLHLR*IRKQAVLLKQPRGPWRYSEQTVSVSSI SRMVL GGPSLSLSKREDYWRLKRGDLHLR*IRKQAVLLKQPRGPWRYSEQTVSVSSI SRMVL
yej0 wildtype VQT 660 Consensus VQT yej0 mutated VQT 660	601	CCWKTAEVCELKKM TSLI IPL*IVAAYWRLWMAGL*LALIKKQAE N*LSQRM RWK*V CCWKTAEVCELKKM TSLI IPL*IVAAYWRLWMAGL*LALIKKQAE N*LSQRM RWK*V
yej0 wildtype YMI 720 Consensus YMI yej0 mutated YMI 720	661	VKANLV*KMVCQKIMNWMVPGSLLWRTRRPLILSLI SMPLCNRWERI LVRKCRQMR VKANLV*KMVCQKIMNWMVPGSLLWRTRRPLILSLI SMPLCNRWERI LVRKCRQMR
yej0 wildtype PQC 780 Consensus PQC yej0 mutated PQC 780	721	SVDHIRMEVSRI PQKPSLKIWLSTMAALTSGLAQW LTFQSEGMM AFLRS*SRK*IMH SVDHIRMEVSRI PQKPSLKIWLSTMAALTSGLAQW LTFQSEGMM AFLRS*SRK*IMH
yej0 wildtype QM* 840 Consensus QM* yej0 mutated QM* 840	781	WWVR*WFLRALLLERMVPWI PAKRTFRSKIAYG PSLPI SLRRTKTPSST*PTLRCLT WWVR*WFLRALLLERMVPWI PAKRTFRSKIAYG PSLPI SLRRTKTPSST*PTLRCLT
yej0 wildtype KYS 900 Consensus KYS yej0 mutated KYS 900	841	L*WMSQ*LVHQ*RQVRKISLR*PPIPCRETAFI CVPIWLI IRAISS TSPVRQQVIS L*WMSQ*LVHQ*RQVRKISLR*PPIPCRETAFI CVPIWLI IRAISS TSPVRQQVIS
yej0 wildtype ARK 960 Consensus ARK yej0 mutated ARK 960	901	*RTPV PARQQEIALHW*QRAAVMLHLRWAMPEALLI SVRMNI PCWIMATIAGVWQRI *RTPV PARQQEIALHW*QRAAVMLHLRWAMPEALLI SVRMNI PCWIMATIAGVWQRI
yej0 wildtype ELV 1020 Consensus ELV yej0 mutated ELV 1020	961	LPLQPLMC*IWRPHNRWYLMQNWT PCVSVLVA*KALVTIRRCGVRQLTPATT*PLMR LPLQPLMC*IWRPHNRWYLMQNWT PCVSVLVA*KALVTIRRCGVRQLTPATT*PLMR
yej0 wildtype SIR 1080 Consensus SIR yej0 mutated SIR 1080	1021	LSKH*RA*RSVSI AVSPVKKAVQFAA*SLVTLILILVLIAAAKVISIAIPWGLMPVG LSKH*RA*RSVSI AVSPVKKAVQFAA*SLVTLILILVLIAAAKVISIAIPWGLMPVG
yej0 wildtype WPL 1140 Consensus	1081	TVPMLMGW*KLTVLPTPSMAR*VMGQQRLAITIVTARVLM L RAGSVGLTDCGVLDPI TVPMLMGW*KLTVLPTPSMAR*VMGQQRLAITIVTARVLM L RAGSVGLTDCGVLDPI

WPL	
yejO mutated	1081 TVPMLMGW*KLTIVLPTPSMAR*VMGQQRLAITIVTARVLMMLRAGSVGLIDCGVLDPI
WPL 1140	
yejO wildtype	1141 PALPQMVRITTRYQTACARMWEIPGYALKRERR*AITWTCKTVRRWNPG*KRPCVRN
TPI 1200	
Consensus	PALPQMVRITTRYQTACARMWEIPGYALKRERR*AITWTCKTVRRWNPG*KRPCVRN
TPI	
yejO mutated	1141 PALPQMVRITTRYQTACARMWEIPGYALKRERR*AITWTCKTVRRWNPG*KRPCVRN
TPI 1200	
yejO wildtype	1201 LTR*KLMTMANLIMMWLEPVAFIRLV*GHRLPRR*AVICQSAMAMAQG*NRRGILRR
VWS 1260	
Consensus	LTR*KLMTMANLIMMWLEPVAFIRLV*GHRLPRR*AVICQSAMAMAQG*NRRGILRR
VWS	
yejO mutated	1201 LTR*KLMTMANLIMMWLEPVAFIRLV*GHRLPRR*AVICQSAMAMAQG*NRRGILRR
VWS 1260	
yejO wildtype	1261 GRS 1263
Consensus	GRS
yejO mutated	1261 GRS 1263

Figure. 6

>yejO Nucleotide alignment Alignment of 2 sequences: yejO wildtype, yejO mutated

Score = 18940.974, Identities = 3790/3791 (99%), Positives = 3790/3791 (99%), Gaps = 0/3791 (0%)

```

yejO wildtype      1 ATGCATCAATCTGGTTCTGTTTCTCTTTGTCGTTCCGCAATATCTGTTCTGGTGGCTA
CA      60
Consensus          ATGCATCAATCTGGTTCTGTTTCTCTTTGTCGTTCCGCAATATCTGTTCTGGTGGCTA
CA
yejO mutated      1 ATGCATCAATCTGGTTCTGTTTCTCTTTGTCGTTCCGCAATATCTGTTCTGGTGGCTA
CA      60

yejO wildtype     61 GCGTTAAGGAAGGTGCGAACAAGTCCCTGATATGAGATCATGTTTGTTCATCTGGAGCC
AT     120
Consensus          GCGTTA GGAAGGTGCGAACAAGTCCCTGATATGAGATCATGTTTGTTCATCTGGAGCC
AT
yejO mutated     61 GCGTTAAGGAAGGTGCGAACAAGTCCCTGATATGAGATCATGTTTGTTCATCTGGAGCC
AT     120

yejO wildtype     121 AGAACAGGGTTCATCATGAGTCATCAACTTACCTTCGCCGACAGTGAATTCAGCAGTA
AG     180
Consensus          AGAACAGGGTTCATCATGAGTCATCAACTTACCTTCGCCGACAGTGAATTCAGCAGTA
AG
yejO mutated     121 AGAACAGGGTTCATCATGAGTCATCAACTTACCTTCGCCGACAGTGAATTCAGCAGTA
AG     180

yejO wildtype     181 CGCCGTCAGACCAGAAAAGAGATTTTCTTGTCCCAGCATGGAGCAGATTCTGCCATGGC
AA     240
Consensus          CGCCGTCAGACCAGAAAAGAGATTTTCTTGTCCCAGCATGGAGCAGATTCTGCCATGGC
AA
yejO mutated     181 CGCCGTCAGACCAGAAAAGAGATTTTCTTGTCCCAGCATGGAGCAGATTCTGCCATGGC
AA     240

yejO wildtype     241 AACATGGTGGAAGTCATCGAGCCGTTTTACCCCAAGGCTGGTAATGGCCGGCGACCTT
AT     300
Consensus          AACATGGTGGAAGTCATCGAGCCGTTTTACCCCAAGGCTGGTAATGGCCGGCGACCTT
AT
yejO mutated     241 AACATGGTGGAAGTCATCGAGCCGTTTTACCCCAAGGCTGGTAATGGCCGGCGACCTT
AT     300

yejO wildtype     301 CCGCTGAAAACCATGCTACGCATTCACTGCATGCAGCATTGGTACAACCTGAGCGATG
GC     360
Consensus          CCGCTGAAAACCATGCTACGCATTCACTGCATGCAGCATTGGTACAACCTGAGCGATGG
C
yejO mutated     301 CCGCTGAAAACCATGCTACGCATTCACTGCATGCAGCATTGGTACAACCTGAGCGATG
GC     360

yejO wildtype     361 GCGATGGAAGATGCTCTGTACGAAATCGCCTCCATGCGTCTGTTTGCCCGTTATCCC
TG     420
Consensus          GCGATGGAAGATGCTCTGTACGAAATCGCCTCCATGCGTCTGTTTGCCCGTTATCCC
TG
yejO mutated     361 GCGATGGAAGATGCTCTGTACGAAATCGCCTCCATGCGTCTGTTTGCCCGTTATCCC
TG     420

yejO wildtype     421 GATAGCGCCTTGCCGGACCGCACCACCATCATGAATTTCCGCCACCTGCTGGAGCAGC
AT     480
Consensus          GATAGCGCCTTGCCGGACCGCACCACCATCATGAATTTCCGCCACCTGCTGGAGCAGC
AT
yejO mutated     421 GATAGCGCCTTGCCGGACCGCACCACCATCATGAATTTCCGCCACCTGCTGGAGCAGC
AT     480

yejO wildtype     481 CAACTGGCCCGCCAATTGTTCAAGACCATCAATCGCTGGCTGGCCGAAGCAGGCGTCA
TG     540
Consensus          CAACTGGCCCGCCAATTGTTCAAGACCATCAATCGCTGGCTGGCCGAAGCAGGCGTCA
TG
yejO mutated     481 CAACTGGCCCGCCAATTGTTCAAGACCATCAATCGCTGGCTGGCCGAAGCAGGCGTCA

```

TG 540	
yej0 wildtype AC 600 Consensus AC	541 ATGACTCAAGGCACCTTGGTCGATGCCACCATCATTGAGGCACCCAGCTCGACCAAGA ATGACTCAAGGCACCTTGGTCGATGCCACCATCATTGAGGCACCCAGCTCGACCAAGA
yej0 mutated AC 600	541 ATGACTCAAGGCACCTTGGTCGATGCCACCATCATTGAGGCACCCAGCTCGACCAAGA
yej0 wildtype TT 660 Consensus TT	601 AAAGAGCAGCAACGCGATCCGGAGATGCATCAGACCAAGAAAGGCAATCAGTGGCACT AAAGAGCAGCAACGCGATCCGGAGATGCATCAGACCAAGAAAGGCAATCAGTGGCACT
yej0 mutated TT 660	601 AAAGAGCAGCAACGCGATCCGGAGATGCATCAGACCAAGAAAGGCAATCAGTGGCACT
yej0 wildtype CC 720 Consensus CC	661 GGCATGAAGGCCACACATTGGTGTGATGCCAAGAGTGGCCTGACCCACAGCCTGGTCA GGCATGAAGGCCACACATTGGTGTGATGCCAAGAGTGGCCTGACCCACAGCCTGGTCA
yej0 mutated CC 720	661 GGCATGAAGGCCACACATTGGTGTGATGCCAAGAGTGGCCTGACCCACAGCCTGGTCA
yej0 wildtype AA 780 Consensus AA	721 ACCGCGGCCAACGAGCATGACCTCAATCAGCTGGGTAATCTGCTGCATGGAGAGGAGC ACCGCGGCCAACGAGCATGACCTCAATCAGCTGGGTAATCTGCTGCATGGAGAGGAGC
yej0 mutated AA 780	721 ACCGCGGCCAACGAGCATGACCTCAATCAGCTGGGTAATCTGCTGCATGGAGAGGAGC
yej0 wildtype TG 840 Consensus TG	781 TTTGTCTCAGCCGATGCCGGCTACCAAGGGGCGCCACAGCGGAGGAGCTGGCCGAGG TTTGTCTCAGCCGATGCCGGCTACCAAGGGGCGCCACAGCGGAGGAGCTGGCCGAGG
yej0 mutated TG 840	781 TTTGTCTCAGCCGATGCCGGCTACCAAGGGGCGCCACAGCGGAGGAGCTGGCCGAGG
yej0 wildtype CA 900 Consensus CA	841 GATGTGGACTGGCTGATCGCCGAGCGCCCCGGCAAGGTAAGAACCTTGAAAACAGCATC GATGTGGACTGGCTGATCGCCGAGCGCCCCGGCAAGGTAAGAACCTTGAAAACAGCATC
yej0 mutated CA 900	841 GATGTGGACTGGCTGATCGCCGAGCGCCCCGGCAAGGTAAGAACCTTGAAAACAGCATC
yej0 wildtype TG 960 Consensus TG	901 CGCAAGAACAAAACGGCCATCAACATCGAATACATGAAAGCCAGCATCCGGGCCAGGG CGCAAGAACAAAACGGCCATCAACATCGAATACATGAAAGCCAGCATCCGGGCCAGGG
yej0 mutated TG 960	901 CGCAAGAACAAAACGGCCATCAACATCGAATACATGAAAGCCAGCATCCGGGCCAGGG
yej0 wildtype GG 1020 Consensus GG	961 GAGCACCCATTTTCGCATCATCAAGCGACAGTTTCGGCTTCGTGAAAGCCAGATACAAGG GAGCACCCATTTTCGCATCATCAAGCGACAGTTTCGGCTTCGTGAAAGCCAGATACAAGG
yej0 mutated GG 1020	961 GAGCACCCATTTTCGCATCATCAAGCGACAGTTTCGGCTTCGTGAAAGCCAGATACAAGG
yej0 wildtype CG 1080 Consensus CG	1021 TTGCTGAAAAACGATAACCAACTGGCGATGTTATTACGCTGGCCAACCTGTTTCGGG TTGCTGAAAAACGATAACCAACTGGCGATGTTATTACGCTGGCCAACCTGTTTCGGG
yej0 mutated CG 1080	1021 TTGCTGAAAAACGATAACCAACTGGCGATGTTATTACGCTGGCCAACCTGTTTCGGG
yej0 wildtype TG 1140 Consensus TG	1081 GACCAAATGATACGTGAGTGGGAGAGATCTCACTAAAAACTGGGGATAACGCCTTAAA GACCAAATGATACGTGAGTGGGAGAGATCTCACTAAAAACTGGGGATAACGCCTTAAA
yej0 mutated TG 1140	1081 GACCAAATGATACGTGAGTGGGAGAGATCTCACTAAAAACTGGGGATAACGCCTTAAA

yej0 wildtype AA 1200 Consensus AA	1141	GCGAAGAAACGGTCTAAATAGGCTGATTC AAGGCATTTACGGGAGAAAAATCGGCTC GCGAAGAAACGGTCTAAATAGGCTGATTC AAGGCATTTACGGGAGAAAAATCGGCTC
yej0 mutated AA 1200	1141	GCGAAGAAACGGTCTAAATAGGCTGATTC AAGGCATTTACGGGAGAAAAATCGGCTC
yej0 wildtype TT 1260 Consensus TT	1201	ACATGAAGAAATGAAATGACTGAGTCAGCCGAGAAGAATTTCCCCGCTTATTTCGCACC ACATGAAGAAATGAAATGACTGAGTCAGCCGAGAAGAATTTCCCCGCTTATTTCGCACC
yej0 mutated TT 1260	1201	ACATGAAGAAATGAAATGACTGAGTCAGCCGAGAAGAATTTCCCCGCTTATTTCGCACC
yej0 wildtype TG 1320 Consensus TG	1261	CCTTATATTCACCCATAGCATTTGGCATCAACTGTTGAGTATGGTGAGACAGTTGATGG CCTTATATTCACCCATAGCATTTGGCATCAACTGTTGAGTATGGTGAGACAGTTGATGG
yej0 mutated TG 1320	1261	CCTTATATTCACCCATAGCATTTGGCATCAACTGTTGAGTATGGTGAGACAGTTGATGG
yej0 wildtype TC 1380 Consensus TC	1321	TTGTCCTGGAAAAAGATATCCAGCTGGTTTATGGGACCGCCAATAATACGAAAAATCAA TTGTCCTGGAAAAAGATATCCAGCTGGTTTATGGGACCGCCAATAATACGAAAAATCAA
yej0 mutated TC 1380	1321	TTGTCCTGGAAAAAGATATCCAGCTGGTTTATGGGACCGCCAATAATACGAAAAATCAA
yej0 wildtype AG 1440 Consensus AG	1381	CTGGCGGAGAACAGCATATAAAAAGAATTTGGTGTAAGTAATAATACTGAAATTAACGG CTGGCGGAGAACAGCATATAAAAAGAATTTGGTGTAAGTAATAATACTGAAATTAACGG
yej0 mutated AG 1440	1381	CTGGCGGAGAACAGCATATAAAAAGAATTTGGTGTAAGTAATAATACTGAAATTAACGG
yej0 wildtype TC 1500 Consensus TC	1441	GGTATCAGTACATTGAAATGAATGGCGCCGAGAATACTCAGTATTAATGACGGTTA GGTATCAGTACATTGAAATGAATGGCGCCGAGAATACTCAGTATTAATGACGGTTA
yej0 mutated TC 1500	1441	GGTATCAGTACATTGAAATGAATGGCGCCGAGAATACTCAGTATTAATGACGGTTA
yej0 wildtype GG 1560 Consensus GG	1501	AAATTGTTCAAATGGGTGGCGGGCAAACCAGACTACGCTCAATAATGGTGTGCTACA AAATTGTTCAAATGGGTGGCGGGCAAACCAGACTACGCTCAATAATGGTGTGCTACA
yej0 mutated GG 1560	1501	AAATTGTTCAAATGGGTGGCGGGCAAACCAGACTACGCTCAATAATGGTGTGCTACA
yej0 wildtype TG 1620 Consensus TG	1561	TTTATGGCGCAGCGAATGATACCACGATTAAAGGCGGGCGCTTAATCGTTGAAAAAGA TTTATGGCGCAGCGAATGATACCACGATTAAAGGCGGGCGCTTAATCGTTGAAAAAGA
yej0 mutated TG 1620	1561	TTTATGGCGCAGCGAATGATACCACGATTAAAGGCGGGCGCTTAATCGTTGAAAAAGA
yej0 wildtype AT 1680 Consensus AT	1621	GGGGGGCCGTCCTTTGTTCGCTATCGAAAAGGGAGGACTACTGGAGGTTAAAGAGGGGGG GGGGGGCCGTCCTTTGTTCGCTATCGAAAAGGGAGGACTACTGGAGGTTAAAGAGGGGGG
yej0 mutated AT 1680	1621	GGGGGGCCGTCCTTTGTTCGCTATCGAAAAGGGAGGACTACTGGAGGTTAAAGAGGGGGG
yej0 wildtype GG 1740 Consensus GG	1681	TTGCATTTGCGGTAGATCAGAAAGCAGGCGGTGCTATTA AAAACAACCACGCGGGCCAT TTGCATTTGCGGTAGATCAGAAAGCAGGCGGTGCTATTA AAAACAACCACGCGGGCCAT
yej0 mutated GG 1740	1681	TTGCATTTGCGGTAGATCAGAAAGCAGGCGGTGCTATTA AAAACAACCACGCGGGCCAT
yej0 wildtype TA 1800	1741	AGGTATTCGGAACAAACCGTCTCGGTCAGTTCGATATCAAGAATGGTATTGCTAATAAA

Consensus TA		AGGTATTCGGAACAAACCGTCTCGGTCAGTTCGATATCAAGAATGGTATTGCTAATAA
yej0 mutated TA 1800	1741	AGGTATTCGGAACAAACCGTCTCGGTCAGTTCGATATCAAGAATGGTATTGCTAATAA
yej0 wildtype CA 1860	1801	TGTTGTTGGAAAACGGCCGGAAGTTTTCGAGTTGAAGAAAATGACTTCGCTTATAAATAC
Consensus CA		TGTTGTTGGAAAACGGCCGGAAGTTTTCGAGTTGAAGAAAATGACTTCGCTTATAAATAC
yej0 mutated CA 1860	1801	TGTTGTTGGAAAACGGCCGGAAGTTTTCGAGTTGAAGAAAATGACTTCGCTTATAAATAC
yej0 wildtype TA 1920	1861	CTGTAGATAGTGGCGGCTTACTGGAGGTTATGGATGGCGGGACTGTAACCTGGCGTTGA
Consensus TA		CTGTAGATAGTGGCGGCTTACTGGAGGTTATGGATGGCGGGACTGTAACCTGGCGTTGA
yej0 mutated TA 1920	1861	CTGTAGATAGTGGCGGCTTACTGGAGGTTATGGATGGCGGGACTGTAACCTGGCGTTGA
yej0 wildtype CA 1980	1921	AAAAAGCAGGCGGAAAATTAATTGTCTCAACGAATGCGCTGGAAGTGAGTGGTCCAAA
Consensus CA		AAAAAGCAGGCGGAAAATTAATTGTCTCAACGAATGCGCTGGAAGTGAGTGGTCCAAA
yej0 mutated CA 1980	1921	AAAAAGCAGGCGGAAAATTAATTGTCTCAACGAATGCGCTGGAAGTGAGTGGTCCAAA
yej0 wildtype TT 2040	1981	GTAAAGGCCAATTTAGTATAAAAGATGGTGTGTCAAAAAATTATGAACTGGATGATGG
Consensus TT		GTAAAGGCCAATTTAGTATAAAAGATGGTGTGTCAAAAAATTATGAACTGGATGATGG
yej0 mutated TT 2040	1981	GTAAAGGCCAATTTAGTATAAAAGATGGTGTGTCAAAAAATTATGAACTGGATGATGG
yej0 wildtype CA 2100	2041	CCGGGCTCATTGTTATGGAGGACACGCAGGCCATTGATACTATCCTTGATAAGCATGC
Consensus CA		CCGGGCTCATTGTTATGGAGGACACGCAGGCCATTGATACTATCCTTGATAAGCATGC
yej0 mutated CA 2100	2041	CCGGGCTCATTGTTATGGAGGACACGCAGGCCATTGATACTATCCTTGATAAGCATGC
yej0 wildtype TC 2160	2101	CTATGCAATCGCTGGGAAAGGATACTGGTACGAAAGTGCAGGCAAATGCCGTATATGA
Consensus TC		CTATGCAATCGCTGGGAAAGGATACTGGTACGAAAGTGCAGGCAAATGCCGTATATGA
yej0 mutated TC 2160	2101	CTATGCAATCGCTGGGAAAGGATACTGGTACGAAAGTGCAGGCAAATGCCGTATATGA
yej0 wildtype TA 2220	2161	TGGTTCGATCATATCAGAAATGGAAGTATCACGTATTCCTCAAAGCCATCTCTGAAAA
Consensus TA		TGGTTCGATCATATCAGAAATGGAAGTATCACGTATTCCTCAAAGCCATCTCTGAAAA
yej0 mutated TA 2220	2161	TGGTTCGATCATATCAGAAATGGAAGTATCACGTATTCCTCAAAGCCATCTCTGAAAA
yej0 wildtype CA 2280	2221	TGGTTATCAACAATGGCCCGCTAACGCTCTGGGCTGGCACAATGGTTAACGTTTCAGT
Consensus CA		TGGTTATCAACAATGGCCCGCTAACGCTCTGGGCTGGCACAATGGTTAACGTTTCAGT
yej0 mutated CA 2280	2221	TGGTTATCAACAATGGCCCGCTAACGCTCTGGGCTGGCACAATGGTTAACGTTTCAGT
yej0 wildtype GT 2340	2281	GAGGGAATGATGGCATTCTTGAGGTCATGAAGCCGCAAATAAATTATGCACCCGCAAT
Consensus GT		GAGGGAATGATGGCATTCTTGAGGTCATGAAGCCGCAAATAAATTATGCACCCGCAAT
yej0 mutated GT 2340	2281	GAGGGAATGATGGCATTCTTGAGGTCATGAAGCCGCAAATAAATTATGCACCCGCAAT
yej0 wildtype TA 2400	2341	TGGTGGGTAAGGTAGTGGTTTCTGAGGGCGCTTCTTTTAGAACGCATGGTGCCGTGGA
Consensus TA		TGGTGGGTAAGGTAGTGGTTTCTGAGGGCGCTTCTTTTAGAACGCATGGTGCCGTGGA

yejO mutated TA 2400	2341	TGGTGGGTAAGGTAGTGGTTTCTGAGGGCGCTTCTTTTAGAACGCATGGTGCCGTGGA
yejO wildtype TA 2460 Consensus TA	2401	CCAGCAAAGCGGACGTTTCGCTCGAAAATAGCGTATGGACCATCATTGCCGATATCAC CCAGCAAAGCGGACGTTTCGCTCGAAAATAGCGTATGGACCATCATTGCCGATATCAC
yejO mutated TA 2460	2401	CCAGCAAAGCGGACGTTTCGCTCGAAAATAGCGTATGGACCATCATTGCCGATATCAC
yejO wildtype GA 2520 Consensus GA	2461	CGACGAACCAAAAACACCCTCCTCAACTTAGCCAACCTTGGCATGTCTGACGCAAATGT CGACGAACCAAAAACACCCTCCTCAACTTAGCCAACCTTGGCATGTCTGACGCAAATGT
yejO mutated GA 2520	2461	CGACGAACCAAAAACACCCTCCTCAACTTAGCCAACCTTGGCATGTCTGACGCAAATGT
yejO wildtype TA 2580 Consensus TA	2521	TTATGATGGATGAGCCAGTGACTCGTTTCATCAGTGACGGCAAGTGCGGAAAATTTTCAT TTATGATGGATGAGCCAGTGACTCGTTTCATCAGTGACGGCAAGTGCGGAAAATTTTCAT
yejO mutated TA 2580	2521	TTATGATGGATGAGCCAGTGACTCGTTTCATCAGTGACGGCAAGTGCGGAAAATTTTCAT
yejO wildtype TA 2640 Consensus TA	2581	CGTTGACCACCAATACCCTGTGGGAAACGGCAATTTTTATATGCGTACCGATATGGC CGTTGACCACCAATACCCTGTGGGAAACGGCAATTTTTATATGCGTACCGATATGGC
yejO mutated TA 2640	2581	CGTTGACCACCAATACCCTGTGGGAAACGGCAATTTTTATATGCGTACCGATATGGC
yejO wildtype CG 2700 Consensus CG	2641	ATCATCAGAGCGATCAGCTCAACGTCACCGGTCAGGCAACAGGTGATTTCAAATATT ATCATCAGAGCGATCAGCTCAACGTCACCGGTCAGGCAACAGGTGATTTCAAATATT
yejO mutated CG 2700	2641	ATCATCAGAGCGATCAGCTCAACGTCACCGGTCAGGCAACAGGTGATTTCAAATATT
yejO wildtype CG 2760 Consensus CG	2701	TGACGGACACCGGTGCCAGCCCGGCAGCAGGAGATAGCCTTACACTGGTAAACAACGGG TGACGGACACCGGTGCCAGCCCGGCAGCAGGAGATAGCCTTACACTGGTAAACAACGGG
yejO mutated CG 2760	2701	TGACGGACACCGGTGCCAGCCCGGCAGCAGGAGATAGCCTTACACTGGTAAACAACGGG
yejO wildtype TG 2820 Consensus TG	2761	GCGGTGATGCTGCATTTACGTTGGGCAATGCCGGAGGCGTTGTTGATATCGGTACGTA GCGGTGATGCTGCATTTACGTTGGGCAATGCCGGAGGCGTTGTTGATATCGGTACGTA
yejO mutated TG 2820	2761	GCGGTGATGCTGCATTTACGTTGGGCAATGCCGGAGGCGTTGTTGATATCGGTACGTA
yejO wildtype AA 2880 Consensus AA	2821	AATATACCTTGCTGGATAATGGCAACCATAGCTGGAGTCTGGCAGAGAAATCGCGCGCA AATATACCTTGCTGGATAATGGCAACCATAGCTGGAGTCTGGCAGAGAAATCGCGCGCA
yejO mutated AA 2880	2821	AATATACCTTGCTGGATAATGGCAACCATAGCTGGAGTCTGGCAGAGAAATCGCGCGCA
yejO wildtype TG 2940 Consensus TG	2881	TTACCCCTTCAACCACTGATGTGCTGAATATGGCGGCCGCACAACCGCTGGTATTTGA TTACCCCTTCAACCACTGATGTGCTGAATATGGCGGCCGCACAACCGCTGGTATTTGA
yejO mutated TG 2940	2881	TTACCCCTTCAACCACTGATGTGCTGAATATGGCGGCCGCACAACCGCTGGTATTTGA
yejO wildtype GG 3000 Consensus GG	2941	CAGAACTGGACACCGTGCGTGAGCGTCTTGGTAGCGTAAAAGGCGTTAGTTACGATAC CAGAACTGGACACCGTGCGTGAGCGTCTTGGTAGCGTAAAAGGCGTTAGTTACGATAC
yejO mutated GG 3000	2941	CAGAACTGGACACCGTGCGTGAGCGTCTTGGTAGCGTAAAAGGCGTTAGTTACGATAC

yej0 wildtype TT 3060 Consensus TT	3001	CGATGTGGAGTTCGGCAATTAACACCCGCAACAACGTGACCACTGATGCGGGAGCTGG CGATGTGGAGTTCGGCAATTAACACCCGCAACAACGTGACCACTGATGCGGGAGCTGG
yej0 mutated TT 3060	3001	CGATGTGGAGTTCGGCAATTAACACCCGCAACAACGTGACCACTGATGCGGGAGCTGG
yej0 wildtype AA 3120 Consensus AA	3061	TTGAGCAAACATTGACGGGCCTGACGCTCGGTATCGATAGCCGTTTCTCCCGTGAAGA TTGAGCAAACATTGACGGGCCTGACGCTCGGTATCGATAGCCGTTTCTCCCGTGAAGA
yej0 mutated AA 3120	3061	TTGAGCAAACATTGACGGGCCTGACGCTCGGTATCGATAGCCGTTTCTCCCGTGAAGA
yej0 wildtype CG 3180 Consensus CG	3121	GCAGTACAATTCGCGGCTTGATCTTTGGTTACTCTCATTCTGATATTGGTTTTGATCG GCAGTACAATTCGCGGCTTGATCTTTGGTTACTCTCATTCTGATATTGGTTTTGATCG
yej0 mutated CG 3180	3121	GCAGTACAATTCGCGGCTTGATCTTTGGTTACTCTCATTCTGATATTGGTTTTGATCG
yej0 wildtype GA 3240 Consensus GA	3181	GCGGCAAAGGTAATATCGATAGCTATAACCCCTGGGGGCTTATGCCGGTTGGGAGCATCA GCGGCAAAGGTAATATCGATAGCTATAACCCCTGGGGGCTTATGCCGGTTGGGAGCATCA
yej0 mutated GA 3240	3181	GCGGCAAAGGTAATATCGATAGCTATAACCCCTGGGGGCTTATGCCGGTTGGGAGCATCA
yej0 wildtype CA 3300 Consensus CA	3241	ACGGTGCCTATGTTGATGGGGTGGTGAAAGTTGACCGTTTTGCCAACACCATCCATGG ACGGTGCCTATGTTGATGGGGTGGTGAAAGTTGACCGTTTTGCCAACACCATCCATGG
yej0 mutated CA 3300	3241	ACGGTGCCTATGTTGATGGGGTGGTGAAAGTTGACCGTTTTGCCAACACCATCCATGG
yej0 wildtype TG 3360 Consensus TG	3301	AGATGAGTAATGGGGCAACAGCGTTTGGCGATTACAATAGTAACGGCGCGGGTGCTCA AGATGAGTAATGGGGCAACAGCGTTTGGCGATTACAATAGTAACGGCGCGGGTGCTCA
yej0 mutated TG 3360	3301	AGATGAGTAATGGGGCAACAGCGTTTGGCGATTACAATAGTAACGGCGCGGGTGCTCA
yej0 wildtype TA 3420 Consensus TA	3361	TTGAGAGCGGGTCCGTTGGGTTGACGGATTGTGGAGTGTAGACCCATCTGGCCTT TTGAGAGCGGGTCCGTTGGGTTGACGGATTGTGGAGTGTAGACCCATCTGGCCTT
yej0 mutated TA 3420	3361	TTGAGAGCGGGTCCGTTGGGTTGACGGATTGTGGAGTGTAGACCCATCTGGCCTT
yej0 wildtype GG 3480 Consensus GG	3421	CCGGCTTTACCACAGATGGTCAGGACTACACGTTATCAAACGGCATGCGCGGGATGT CCGGCTTTACCACAGATGGTCAGGACTACACGTTATCAAACGGCATGCGCGGGATGT
yej0 mutated GG 3480	3421	CCGGCTTTACCACAGATGGTCAGGACTACACGTTATCAAACGGCATGCGCGGGATGT
yej0 wildtype GC 3540 Consensus GC	3481	GAAATACCCGGATATTACGCGCTGAAGCGGGAACGGCGGTAAGCTATCACATGGACCT GAAATACCCGGATATTACGCGCTGAAGCGGGAACGGCGGTAAGCTATCACATGGACCT
yej0 mutated GC 3540	3481	GAAATACCCGGATATTACGCGCTGAAGCGGGAACGGCGGTAAGCTATCACATGGACCT
yej0 wildtype TT 3600 Consensus TT	3541	AAAACGGTACGACGCTGGAACCCTGGCTGAAAGCGGCCGTGCGTCAGGAATACGCCGA AAAACGGTACGACGCTGGAACCCTGGCTGAAAGCGGCCGTGCGTCAGGAATACGCCGA
yej0 mutated TT 3600	3541	AAAACGGTACGACGCTGGAACCCTGGCTGAAAGCGGCCGTGCGTCAGGAATACGCCGA
yej0 wildtype	3601	CTAACCAGGTGAAAGTTAATGACGATGGCAAATTTAATAATGATGTGGCTGGAACCAG

TG 3660	
Consensus	CTAACCAGGTGAAAGTTAATGACGATGGCAAATTTAATAATGATGTGGCTGGAACCAG
TG	
yejO mutated	3601 CTAACCAGGTGAAAGTTAATGACGATGGCAAATTTAATAATGATGTGGCTGGAACCAG
TG 3660	
yejO wildtype	3661 GCGTTTATCAGGCTGGTATAAAGGTCATCGTTTACCCCGACGTTAAGCGGTCATTTGTC
AG 3720	
Consensus	GCGTTTATCAGGCTGGTATAAAGGTCATCGTTTACCCCGACGTTAAGCGGTCATTTGTC
AG	
yejO mutated	3661 GCGTTTATCAGGCTGGTATAAAGGTCATCGTTTACCCCGACGTTAAGCGGTCATTTGTC
AG 3720	
yejO wildtype	3721 TCAGCTATGGCAATGGCGCAGGGGTAGAATCGCCGTGGAATACTCAGGCGGGTGTGGT
CT 3780	
Consensus	TCAGCTATGGCAATGGCGCAGGGGTAGAATCGCCGTGGAATACTCAGGCGGGTGTGGT
CT	
yejO mutated	3721 TCAGCTATGGCAATGGCGCAGGGGTAGAATCGCCGTGGAATACTCAGGCGGGTGTGGT
CT 3780	
yejO wildtype	3781 GGACGTTCTGA 3791
Consensus	GGACGTTCTGA
yejO mutated	3781 GGACGTTCTGA 3791

PHAGE-RESISTANT MICROORGANISMS

FIELD OF THE INVENTION

[0001] The present invention provides genetically modified microorganisms that are resistant to infection by bacteriophages. The present invention also provides a method of making bacteriophage-infection resistant microorganisms.

BACKGROUND OF THE INVENTION

[0002] Bacterial cultures are the center of the biotechnology industry. Whether a process is profitable or not depends on proper growth. Therefore, numerous and rigorous measures are employed to maintain control over culture conditions. However, these cultures are prone to contamination by other microorganisms, or they can be infected by a great number of viruses, called bacteriophages or phages.

[0003] During phage infection, the phage recognize and attack their host cell in the lytic cycle until the host is completely destroyed releasing hundreds of viral particles which has the potential of attacking the remaining sensitive cells in the culture. Prior to the present invention, the threat of phage infection is one of the most serious problems affecting bacterial cultures of biotechnological interest leading to significant losses in the production process. This has led to the need of finding methods that allow for the generation of enhanced strains which are resistant to infection by bacteriophages.

[0004] U.S. Pat. No. 5,240,841A describes a method to generate an *E. coli* strain resistant to bacteriophage Q β is described. This method consists of isolating a specific region of the corresponding viral replicase gene. This region encodes for the peptide moiety of the viral replicase, which has the function of binding the viral genome in a specific sequence for its replication. After being isolated, this moiety can be introduced in the *E. coli* genome. When expressed as a peptide, it competes for the binding site with viral replicas preventing the replication and spreading of the virus, which provides resistance to the host. However, in order to apply this strategy to all bacteriophages which infect *E. coli*, a peptide must be generated for each phage in an *E. coli* strain, which is inviable.

[0005] Hong J et al. (Hong, J. et al. Identification of host receptor and receptor-binding module of a newly sequenced T5-like phage EPS7. FEMS Microbiology letters. Vol 289 (2). pp 202-209. 2008.) demonstrated that the BtuB protein, which is a transmembrane transporter in *E. coli* involved in the transport of vitamin B12, acts as a receptor of T5-like phage EPS7. By making a series of mutations of the encoding gene of this protein, btuB, the infection by phages was blocked demonstrating that this receptor has an important role during the adsorption process of the phage.

[0006] Knirel, Y A. et al. (Knirel, Y A. et al. Variations in O-antigen biosynthesis and O-acetylation associated with altered phage sensitivity in *Escherichia coli* 4s. Journal of Bacteriology. Vol 197(5). pp 905-912. 2015) describes variations in the synthesis and structure of antigen O of *E. coli* strain 4s isolated from fecal matter from horse. These mutations induce resistance to bacteriophage G7C and further modify the interaction of *E. coli* 4s with other different bacteriophages leading to both resistance and sensitivity to the host cell.

[0007] WO1997020917A2 describes the use of a gene called AbiE, which encodes for a protein that interrupts the

infection by phages. This gene resides in native form in the *Lactococcus lactis* strain. This gene was isolated and cloned in plasmid pSRQ800. The transformation of *Lactococcus lactis* or other microorganisms used in the dairy industry gives resistance to infection by phages 936, c2 and P335.

[0008] WO2001007566A2 describes a genetic system capable of imparting resistance to infection by phages, which consists of two plasmids, pCRB33 and pCRB63. After transformation in *Streptococcus thermophilus* strain, this strain acquires resistance to infection by phages. Plasmids encode for elements of a type I methylation-restriction system called "s" subunits. Both plasmids have these incomplete genetic elements. Due to the great homology in the sequence of "s" subunits, they recombine inside the cell producing a third plasmid (pCRB96). This recombination produces a complete ORF for the "S" subunit imparting resistance to phages.

[0009] CA2311598A1 describes a method and elements necessary for imparting resistance to phages in *Lactococcus lactis* and other strains used in dairy industry. This patent describes the use of an Abi900 protein encoded by plasmid pSRQ900. By being transformed by this plasmid in the strain of interest, resistance to phages 936, c2 and P335 by the infection interruption mechanism is provided.

[0010] Denes et al. (Denes, T., et al., Appl Environ Microbiol, 81 (13), pp 4295-4305 (2015)) isolated strains of *Listeria monocytogens* resistant to phages LP-048 and LP-125. By sequencing, they found mutations in two key loci for the adsorption of phages LP-048 and LP-125.

[0011] Proper phage isolation is necessary to corroborate that one has phage-resistant strains. Examples of phage-isolation protocols known in the art include: (1) (Uc-Mass, Augusto. et al. An orthologue of the cor gene is involved in the exclusion of temperature lambdoid phages. Evidence that Cor inactivates FhuA receptor functions. Virology., Vol 329. pp 425-433. 2004.) and (2) (Kameyama, Luis. et al. Characterization of wild lambdoid bacteriophages: detection of a wide distribution of phageimmunity groups and identification of a nus-dependent, nonlambdoid phage group. Virology. Vol 263. pp 100-111. 1999), or they can be acquired from microbial collections, such as the ATCC, etc.

[0012] It is important to note that prior to the present invention no disclosure has been made that relates mutations or deletions in tfaD and yejO genes to impart resistance to different types of phages which infect *E. coli*.

SUMMARY OF THE INVENTION

[0013] The present invention provides a method for generating genetically modified microorganisms that are resistant to infection by different phages.

[0014] The present invention also provides genetically modified microorganisms that are resistant to infection by different phage families.

[0015] Further, the present invention provides *E. coli* strains that have point mutations or deletions in different genes, and which also impart resistance to various phage families.

[0016] The present invention provides *E. coli* strains that are resistant to phages of the Siphoviridae and Myoviridae families.

[0017] The present invention provides *E. coli* strains that are resistant to phages λ , ϕ 80 and T4.

[0018] Moreover, the present invention provides *E. coli* strains that are resistant to phages λ , $\phi 80$ and T4 and that retain their kinetic constants relative to wild type strains.

[0019] The above objects highlight certain aspects of the invention. Additional objects, aspects and embodiments of the invention are found in the following detailed description of the invention.

BRIEF DESCRIPTION OF THE DRAWINGS

[0020] A more complete appreciation of the invention and many of the attendant advantages thereof will be readily obtained as the same becomes better understood by reference to the following Figures in conjunction with the detailed description below.

[0021] FIG. 1 shows the difference between wildtype *E. coli* K-12 and *E. coli* LCT-BF-01 strains infected by phage λ , $\phi 80$ and T4 in M9 agar medium. It was observed that strain LCT-BF-01 grew normally, while the wild type strain exhibited lytic plaques which are indicative of cellular lysis.

[0022] FIG. 2 shows the difference between growth of *E. coli* LCT-BF-01 and wild type *E. coli* K-12 strains in liquid M9 medium before and after infection by phages λ , $\phi 80$ and T4. LCT-BF-01 strain is resistant to infection by bacteriophages, while wild type K12 strain is lysed. Growth kinetics of wild type strains (X) and LCT-BF-01 strain (O). The arrow indicates the point where cultures are infected with the phage mixture.

[0023] FIG. 3 shows a protein alignment of the wildtype TfaD protein (SEQ ID NO: 2) and the mutated TfaD protein (SEQ ID NO: 4).

[0024] FIG. 4 shows a protein alignment of the wildtype TfaD gene (SEQ ID NO: 1) and the mutated TfaD gene (SEQ ID NO: 3).

[0025] FIG. 5 shows a protein alignment of the wildtype YejO protein (SEQ ID NO: 6) and the mutated YejO protein (SEQ ID NO: 8).

[0026] FIG. 6 shows a protein alignment of the wildtype YejO gene (SEQ ID NO: 5) and the mutated YejO gene (SEQ ID NO: 7).

DETAILED DESCRIPTION OF THE INVENTION

[0027] Unless specifically defined, all technical and scientific terms used herein have the same meaning as commonly understood by a skilled artisan in enzymology, biochemistry, cellular biology, molecular biology, and the medical sciences.

[0028] All methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, with suitable methods and materials being described herein. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. Further, the materials, methods, and examples are illustrative only and are not intended to be limiting, unless otherwise specified.

[0029] In order to better understand the object of the present invention, the following definitions and abbreviations are established.

[0030] The terms “gene” or “genes” refer to biological molecules composed of nitrogen compounds or nitrogen bases known in the state of the art, such as Adenine,

Guanine, Cytosine and Thymine. Genes are molecules which transmit information in a cell for the biological synthesis of enzymes.

[0031] The term “locus” refers to the fixed position of a gene on a chromosome.

[0032] The term “loci” refers to the plural of “locus”, i.e., the fixed positions of two or more genes on a chromosome.

[0033] The term “substrate” refers to a molecule which can be used as a carbon source for the microorganism to grow or to be used as a desired product. Examples of substrate can be carbohydrates, lipids, proteins, organic acids, alcohols, aldehydes, ketones, hydrocarbons, etc.

[0034] The term “deletion or removal of genes” refers to the procedure of totally or partially removing a gene, modifying the reading frame or adding a stop codon in any region of the gene other than the natural stop codon; it also refers to adding or removing regions which prevent the transcription and/or reduction of the gene.

[0035] The term “plaque-forming units” refers to cells which were infected by the bacteriophage or to the number of bacteriophages which infected a cell from the culture.

[0036] The terms “phages” or “bacteriophage” refer to a virus which is capable of infecting bacteria and which can produce the cell rupture of bacteria during the infection cycle. Examples of these phages can be M13, T4, Lambda or any other virus which is described in the state of the art which causes infection of bacteria, whether by the lytic cycle or lysogenic cycle. The lytic and lysogenic cycles of a phage are widely known by any person related to the field of the invention.

[0037] The term “biomass” refers to the total amount of organic matter which makes up the culture and which corresponds to a single type of microorganism, in this case, the producing strain and its exponential growth resulting from the fermentation process. Biomass is spectrophotometrically determined by optical density at 600 nm and by dry weight in thermoscale expressed in g/L.

[0038] The term “inoculum” refers to the initial biomass portion corresponding to the strain of interest to initiate the fermentation process.

[0039] The term “fermentation” refers to the catabolic metabolism in which the oxidation of the carbon source can be complete by having Oxygen as final electron acceptor or incomplete, wherein an organic compound is produced which functions as electron donor and electron acceptor at the same time and wherein ATP is produced by phosphorylation at substrate level.

[0040] The term “culture medium” refers to the solution which contains the necessary nutrients to allow the growth of the strain of interest. Known media in the state of the art are M9, LB 2YT, and any other medium which is reported in the state of the art which could be useful for the growth of the strain of interest.

[0041] The term “anaerobic conditions” refers to a fermentation period in which oxygen is fed to the reactor tank, which acts as ultimate electron acceptor and the oxidation of the carbon source is complete.

[0042] The term “expressed” refers to the gene or set of genes which are transcribed in certain conditions during fermentation.

[0043] The term “wild type strain” refers to an organism which retains the original genetic material of its species, i.e., its genetic information has not been modified.

[0044] The term “ μ ” refers to the specific rate of growth of the strain of interest, expressed in h^{-1} , which depends on the concentration of nutrients in the medium and on operating parameters, such as agitation and aeration.

[0045] The term “Qs” refers to the consumption of a specific substrate, expressed in $(\text{g/g}\cdot\text{h})$, i.e., the mass of substrate consumed by biomass unit during a certain period.

[0046] The term “attenuated phage or bacteriophage” refers to a virus whose genome is capable of replicating together with that of its host and does not cause cellular death in a state called lysogeny.

[0047] The term “reactor” refers to a physical space built from a suitable material in which, in a controlled way, a chemical, biochemical or biological reaction can take place, or combinations thereof. Different types of reactors can be found in the state of the art. By way of example, reactors such as a continuous stirred-tank reactors (CSTR), piston flow reactors, fluidized bed reactors and packed bed reactors (PBR) are described. Some of the features of reactors are: a) their resistance to corrosion due to the reaction which is taking place; b) their capacity for monitoring and controlling operation variables, such as temperature, agitation, pH, concentration of dissolved gases, pressure, etc.; c) the operation mode, which can be continuous, semi-continuous or batch (different operation modes in which a reactor can work are described in the state of the art); d) the capacity of using different types of catalysts which will carry out the reaction, for example, the catalysts can be dissolved or trapped or immobilized (different modes in which a catalyst can carry out the reaction inside a reactor are described in the state of the art). The present invention provides a method that allows microorganisms sensitive to infection by phages to acquire resistance to infection by phages, due to genetic changes.

[0048] More specifically, the present invention provides a method that allows microorganisms sensitive to infection by phages to be resistant to infection by one or more phages at the same time.

[0049] Further, the present invention provides a method that allows microorganisms sensitive to infection by phages to be resistant to infection by one or more phages from the same family at the same time.

[0050] Further, the present invention provides a method that allows microorganisms sensitive to infection by phages to be resistant to infection by one or more phages from different families at the same time.

[0051] Moreover, the present invention provides microorganisms that are capable of resisting an infection by one or more types of phages due to mutations in certain genes.

[0052] Further, the present invention provides microorganisms that are capable of resisting an infection by one or more phage families due to genetic changes.

[0053] Finally, the present invention provides microorganisms that are capable of resisting an infection by one or more phage families and also retain the same kinetic constants.

[0054] In another embodiment of the present invention, a method for generating microorganisms resistant to infection by phage λ is provided, wherein the phage λ come into contact with the microorganism during a certain time to allow the infection. Subsequently, the culture is allowed to recover during a certain time. Bacteria which could survive the infection are isolated and biochemically, microbiologically and genetically characterized as follows:

[0055] I. In a flask containing M9 culture medium, the microorganism is allowed to grow until reaching an

optical density from 0.6 to 5, more specifically from 1 to 3 and more specifically from 2 to 2.5.

[0056] II. Phage λ is added to the culture medium, wherein the concentration of the phage is at least 100 plaque-forming units, more specifically at least 250 plaque-forming units, and more specifically at least 500 plaque-forming units.

[0057] III. Phage infection is allowed from 30 minutes to 6 hours, more specifically from 2 hours to 4 hours, and more specifically from 2.5 hours to 3.5 hours at room temperature, more preferably at a temperature ranging from 20 to 25° C.

[0058] IV. The culture is allowed to lyse from 1 hour to 8 hours, more specifically from 3 hours to 6 hours and more specifically from 4 hours to 5 hours at 37° C., more preferably at a temperature ranging from 35 to 39° C.

[0059] V. The culture is allowed to recover from 1 hour to 24 hours, more specifically from 6 hours to 20 hours and more specifically from 10 hours to 16 hours.

[0060] Recovery may be at room temperature, more preferably at a temperature ranging from 20 to 25° C.

[0061] VI. Colonies are isolated into plates with M9 medium or with other medium suitable for cell growth.

[0062] VII. It is verified that the colonies are resistant to phage λ by repeating steps I to VI at least twice.

[0063] In another embodiment of the present invention, a method for generating microorganisms resistant to infection by phage $\phi 80$ is provided, wherein the phage $\phi 80$ come into contact with the microorganism during a certain time to allow the infection. Subsequently, the culture is allowed to recover during a certain time. Bacteria which could survive the infection are isolated and biochemically, microbiologically and genetically characterized as follows:

[0064] I. In a flask containing M9 culture medium, the microorganism is allowed to grow until reaching an optical density from 0.6 to 5, more specifically from 1 to 3 and more specifically from 2 to 2.5

[0065] II. Phage $\phi 80$ is added to the culture medium, wherein the concentration of the phage is at least 100 plaque-forming units, more specifically at least 250 plaque-forming units, and more specifically at least 500 plaque-forming units.

[0066] III. Phage infection is allowed from 30 minutes to 6 hours, more specifically from 2 hours to 4 hours, and more specifically from 2.5 hours to 3.5 hours at room temperature, more preferably at a temperature ranging from 20 to 25° C.

[0067] IV. The culture is allowed to lyse from 1 hour to 8 hours, more specifically from 3 hours to 6 hours and more specifically from 4 hours to 5 hours at 37° C., more preferably at a temperature ranging from 35 to 39° C.

[0068] V. The culture is allowed to recover from 1 hour to 24 hours, more specifically from 6 hours to 20 hours and more specifically from 10 hours to 16 hours. Recovery may be at room temperature, more preferably at a temperature ranging from 20 to 25° C.

[0069] VI. Colonies are isolated into plates with M9 medium or with other medium suitable for cell growth.

[0070] VII. It is verified that the colonies are resistant to phage $\phi 80$ by repeating steps I to VI at least twice.

[0071] In another embodiment of the present invention, a method for generating microorganisms resistant to infection

by phage T4 is provided, wherein the phage T4 come into contact with the microorganism during a certain time to allow the infection. Subsequently, the culture is allowed to recover during a certain time. Bacteria which could survive the infection are isolated and biochemically, microbiologically and genetically characterized as follows:

[0072] I. In a flask containing M9 culture medium, the microorganism is allowed to grow until reaching an optical density from 0.6 to 5, more specifically from 1 to 3 and more specifically from 2 to 2.5.

[0073] II. Phage T4 is added to the culture medium, wherein the concentration of the phage is at least 100 plaque-forming units, more specifically at least 250 plaque-forming units, and more specifically at least 500 plaque-forming units.

[0074] III. Phage infection is allowed from 30 minutes to 6 hours, more specifically from 2 hours to 4 hours, and more specifically from 2.5 hours to 3.5 hours at room temperature, more preferably at a temperature ranging from 20 to 25° C.

[0075] IV. The culture is allowed to lyse from 1 hour to 8 hours, more specifically from 3 hours to 6 hours and more specifically from 4 hours to 5 hours at 37° C., more preferably at a temperature ranging from 35 to 39° C.

[0076] V. The culture is allowed to recover from 1 hour to 24 hours, more specifically from 6 hours to 20 hours and more specifically from 10 hours to 16 hours. Recovery may be at room temperature, more preferably at a temperature ranging from 20 to 25° C.

[0077] VI. Colonies are isolated into plates with M9 medium or with other medium suitable for cell growth.

[0078] VII. It is verified that the colonies are resistant to phage T4 by repeating steps I to VI at least twice.

[0079] In another embodiment of the present invention, a method for generating bacteria from the *E. coli* genus which have mutations in the *tfaD* and *yejO* genes and which are also resistant to infection by phages is provided, wherein the phages come into contact with the microorganism during a certain time to allow the infection. Subsequently, the culture is allowed to recover during a certain time. Bacteria which could survive the infection are isolated and biochemically, microbiologically and genetically characterized as follows:

[0080] I. To an *E. coli* strain, mutations are made in the *tfaD* and *yejO* genes.

[0081] II. In a flask containing M9 culture medium, the *E. coli* strain is allowed to grow with the *tfaD* and *yejO* mutations until reaching an optical density from 0.6 to 5, more specifically 1 to 3 and more specifically 2 to 2.5.

[0082] III. Phages λ , ϕ 80 and T4 and are added to the culture medium, separately or mixed, wherein the concentration of the phage is at least 100 plaque-forming units, more specifically at least 250 plaque-forming units, and more specifically at least 500 plaque-forming units.

[0083] IV. Phage infection is allowed from 30 minutes to 6 hours, more specifically from 2 hours to 4 hours, and more specifically from 2.5 hours to 3.5 hours at room temperature, more preferably at a temperature ranging from 20 to 25° C.

[0084] V. The culture is allowed to lyse from 1 hour to 8 hours, more specifically from 3 hours to 6 hours and

more specifically from 4 hours to 5 hours at 37° C., more preferably at a temperature ranging from 35 to 39° C.

[0085] VI. The culture is allowed to recover from 1 hour to 24 hours, more specifically from 6 hours to 20 hours and more specifically from 10 hours to 16 hours. Recovery may be at room temperature, more preferably at a temperature ranging from 20 to 25° C.

[0086] VII. Colonies are isolated into plates with M9 medium or with other medium suitable for cell growth.

[0087] VIII. It is verified that the colonies are resistant to the phages λ , ϕ 80 and T4, separately or mixed by repeating steps I to VII at least twice.

[0088] The above written description of the invention provides a manner and process of making and using it such that any person skilled in this art is enabled to make and use the same, this enablement being provided in particular for the subject matter of the appended claims, which make up a part of the original description.

[0089] As used herein, the phrases “selected from the group consisting of,” “chosen from,” and the like include mixtures of the specified materials.

[0090] Where a numerical limit or range is stated herein, the endpoints are included. Also, all values and subranges within a numerical limit or range are specifically included as if explicitly written out.

[0091] The above description is presented to enable a person skilled in the art to make and use the invention, and is provided in the context of a particular application and its requirements. Various modifications to the preferred embodiments will be readily apparent to those skilled in the art, and the generic principles defined herein may be applied to other embodiments and applications without departing from the spirit and scope of the invention. Thus, this invention is not intended to be limited to the embodiments shown, but is to be accorded the widest scope consistent with the principles and features disclosed herein.

EXAMPLES

[0092] The following examples are intended to clarify the novelty of the present invention. It should be understood that the following examples do not limit the scope of the present invention. From the disclosure of the invention, as well as the following examples, a person skilled in the field of the invention can make some modifications, which in any way remain within the framework protected by this invention.

Example 1. Generation of Strains Resistant to Phages λ , ϕ 80 and T4

[0093] In the case of the present invention, different environment phages which infect different microorganisms were isolated, although, to exemplify the method of the present invention, the *E. coli* bacteria was used.

[0094] Different culture media were used to make bacteria come into contact with phages. Some of the media were LB agar medium and M9 agar medium (Sambrook, J., and Green, M. (2012). *Molecular cloning: a laboratory manual*, 4th edition. Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.).

[0095] A culture of wild type *E. coli* K-12 in liquid M9 medium was prepared, it was left to grow from 6 hours to 24 hours, after that, a mixture of phages λ , ϕ 80 and T4 was added. A cycle of infection was allowed from 30 minutes to

6 hours at room temperature. Thereafter, the culture was incubated at 30° C. from 1 hour to 8 hours for the culture to clarify, in this moment most of the cells are dead. Subsequently, incubation was continued to allow for the reproduction of phage-resistant cells. After a period from 1 hour to 24 hours, it was observed that the culture starts growing again and it was taken as inoculum to isolate the resistant colonies.

[0096] Plates were seeded by extension in M9 agar. After 16 hours of incubation at 37° C., phage-resistant colonies were observed and were selected to be infected again.

[0097] Selected candidate colonies were challenged against the phage by the pour plate method. First, a culture was left to grow in liquid from the strain of interest for 6 hours, subsequently, it was infected with the phage allowing for an infection cycle to happen for 1 hour at room temperature. Thereafter, infected cells were mixed in soft M9 agar medium and were incubated for 16 h at 37° C. At the end of this incubation period, it was observed that resistant strains did not exhibit lytic plaques, while sensitive strains did (FIG. 1).

[0098] Isolated colonies were subjected to at least 5 infection and isolation cycles. After the infection and isolation cycles, strains were biochemically, microbiologically and genetically reviewed in order to verify that genetic modifications made at the beginning were the cause of the resistance. The strain that resisted infection by the tested phages was named LCT-BF-01.

[0099] In order to demonstrate that strain LCT-BF-01 was resistant to phages λ , $\phi 80$ and T4 and grew in liquid medium, cultures in a 14 L reactor were made using M9 medium at different operating conditions. The operating conditions are shown in Table 1.

TABLE 1

Operating conditions of the reactor.	
Condition	Values
pH	6.8-8
Temperature	20-40° C.
Dissolved oxygen	0.1-8 mg/L

[0100] The reactor was prepared, sterilized at 121° C. and pressure of 15 psig, inoculated with a colony from strain LCT-BF-01 with sterilization and was allowed to grow for six hours. Subsequently, it was infected by a mixture of phages λ , $\phi 80$ and T4 and its growth, pH, temperature and oxygen were further monitored in the reactor. After 16 hours of growth, it was observed that the culture did not clarify in any moment during fermentation (FIG. 2). The same experiment was made with the wild type *E. coli* K-12 strain, this strain did not exhibit growth after infection (FIG. 2). With this example it was demonstrated that strain LCT-BF-01 is resistant to phages λ , $\phi 80$ and T4.

[0101] Subsequently, strain LCT-BF-01 was sequenced in an Illumina MiniSeq System sequencer, using the bacterial sequencing kit and following the manufacturer's instructions (Illumina Inc.), in order to corroborate the generated mutations. In table 2, genes that underwent mutations during the corresponding infection process are shown.

TABLE 2

Identified mutations in different <i>E. coli</i> genes		
Gene	Position of mutation in protein	Aminoacid changed
TfaD	69	A→T
YejO	23	W→R

TfaD wildtype gene appears as SEQ ID NO: 1 and the wildtype TfaD protein appears as SEQ ID NO: 2. The mutated TfaD gene appears as SEQ ID NO: 3 and the corresponding mutated TfaD protein appears as SEQ ID NO: 4.

YejO wildtype gene appears as SEQ ID NO: 5 and the wildtype YejO protein appears as SEQ ID NO: 6. The mutated YejO gene appears as SEQ ID NO: 7 and the corresponding mutated YejO protein appears as SEQ ID NO: 8.

Example 2. Comparison of Kinetic Parameters of Strains LCT-BF-01 and Wild Type *E. coli* K-12 Strain

[0102] Strains LCT-BF-01 and wild type *E. coli* K-12 were cultured in M9 medium in a 14 L reactor in order to determinate the kinetic parameters in aerobic conditions. The culture took 24 h, the operating conditions are shown in Table 3.

TABLE 3

Operating conditions of the reactor.	
Condition	Values
pH	7
Temperature	37° C.
Dissolved oxygen	2 mg/L

[0103] The concentration of glucose and organic acids were monitored during fermentation using an HPLC Ultimate 3000 equipment (Thermo) with an index of refraction detector using a Rezex-ROA organic acids H⁺ column. Results of fermentations are shown in Table 4.

TABLE 4

Kinetic constants of strains LCT-BF-01 and Wild type		
Variable	Strain LCT-BF-01	Wild type (WT)
μ	0.44	0.45
Qs	0.88	0.89

[0104] These results demonstrate that strains developed in the present invention are resistant to phages from different families and have the same kinetic constants as WT, in spite of having mutations of TfaD and YejO genes.

[0105] Numerous modifications and variations on the present invention are possible in light of the above teachings. It is, therefore, to be understood that within the scope of the accompanying claims, the invention may be practiced otherwise than as specifically described herein.

REFERENCES

- [0106] Denes, T., et. al., 2015. Appl Envirom Microbiol, 81 (13), pp 4295-4305.
- [0107] Hong, J. et al. Identification of host receptor and receptor-binding module of a newly sequenced T5-like phage EPS7. FEMS Microbiology letters. Vol 289(2). pp 202-209. 2008.
- [0108] Kameyama, Luis. et al. Characterization of wild lambdoid bacteriophages: detection of a wide distribution of phageimmunity groups and identification of a nus-dependent, nonlambdoid phage group. Virology. Vol 263. pp 100-111. 1999.
- [0109] Knirel, Y A. et al. Variations in O-antigen biosynthesis and O-acetylation associated with altered phage sensitivity in *Escherichia coli* 4s. Journal of Bacteriology. Vol 197(5). pp 905-912. 2015.
- [0110] Sambrook, J., and Green, M. (2012). Molecular cloning: a laboratory manual, 4th edition. Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.
- [0111] Uc-Mass, Augusto. et al. An orthologue of the core gene is involved in the exclusion of temperata lambdoid phages. Evidence that Cor inactivates FhuA receptor functions. Virology., Vol 329. pp 425-433. 2004.
- [0112] Wang, X., Kim, Y., Ma, Q., Hong, S. H., Pokusaeva, K., Sturino, J. M., & Wood, T. K. (2010). Cryptic prophages help bacteria cope with adverse environments. Nature communications, 1, 147).
- [0113] Patent: CA2311598A1
- [0114] Patent: EP2534252B1
- [0115] Patent: WO1997020917A2
- [0116] Patent: WO2001007566A2

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 8

<210> SEQ ID NO 1

<211> LENGTH: 438

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic tfaD wildtype gene

<400> SEQUENCE: 1

```
attccagctg gcttcctggc tgttttcaac agtgatgagt catcgtggca tctcgttgaa    60
gatcatcggg gtaaaacggt ttatgacgta gcgtcagggg acgcgttatt tatttctgaa    120
ctcgggtccgt taccggaaaa gtgtacctgg ttatcgccgg aaggggagtt tcagaagtgg    180
aacggtacag cctgggtgaa agatgcagaa gcagaaaaac tgttccggat tcgggaggcg    240
gaagaaacaa aaaacagcct gatgcaggta gccagtgagc atattgccc acttcaggat    300
gctgtagatc tggaaatcgc aacggaggaa gaaacctcat tgctggaagc ctggaaaaaa    360
tatcgggtgt tgctgaaccg tgttgatata tcaactgcac ctgatattga gtggcctacg    420
aacctgtca gggagtaa                                     438
```

<210> SEQ ID NO 2

<211> LENGTH: 145

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic tfaD wildtype protein

<400> SEQUENCE: 2

```
Ile Pro Ala Gly Phe Val Ala Val Phe Asn Ser Asp Glu Ser Ser Trp
1           5           10           15
His Leu Val Glu Asp His Arg Gly Lys Thr Val Tyr Asp Val Ala Ser
20          25          30
Gly Asp Ala Leu Phe Ile Ser Glu Leu Gly Pro Leu Pro Glu Asn Val
35          40          45
Thr Trp Leu Ser Pro Glu Gly Phe Gln Lys Trp Asn Gly Thr Ala
50          55          60
Trp Val Lys Asp Ala Glu Ala Glu Lys Leu Phe Arg Ile Arg Glu Ala
65          70          75          80
Glu Glu Thr Lys Asn Ser Leu Met Gln Val Ala Ser Glu His Ile Ala
```

-continued

85	90	95	
Pro Leu Gln Asp Ala Val Asp Leu Glu Ile Ala Thr Glu Glu Glu Thr			
100	105	110	
Ser Leu Leu Glu Ala Trp Lys Lys Tyr Arg Val Leu Leu Asn Arg Val			
115	120	125	
Asp Thr Ser Thr Ala Pro Asp Ile Glu Trp Pro Thr Asn Pro Val Arg			
130	135	140	
Glu			
145			
<210> SEQ ID NO 3			
<211> LENGTH: 400			
<212> TYPE: DNA			
<213> ORGANISM: Artificial Sequence			
<220> FEATURE:			
<223> OTHER INFORMATION: Synthetic tfaD mutated gene			
<400> SEQUENCE: 3			
attccagctg gcttcgtggc tgttttcaac agtgatgagt catcgtggca tctcgttgaa			60
gatcatcggg gtaaaacggt ttatgacgtg gcttcgcgcg acgcgttatt tattttctgaa			120
ctcggtcctg taccggaaaa tgttacctgg ttatcgccgg aaggggagtt tcagaagtgg			180
aacggcacag cctgggtgaa ggatacggaa gcagaaaaac tgttccggat ccgggagggc			240
gaagaaacaa aaaacagcct gatgcaggta gccagtgagc atattgccc acttcaggat			300
gctgtagatc tggaatcgc aacggaggaa gaaacctcat tgctggaagc ctggaaaaaa			360
tatcgggtgt tgmtgaaccg tgttgataca tcaactgcac			400
<210> SEQ ID NO 4			
<211> LENGTH: 145			
<212> TYPE: PRT			
<213> ORGANISM: Artificial Sequence			
<220> FEATURE:			
<223> OTHER INFORMATION: Synthetic tfaD mutated protein			
<400> SEQUENCE: 4			
Ile Pro Ala Gly Phe Val Ala Val Phe Asn Ser Asp Glu Ser Ser Trp			
1	5	10	15
His Leu Val Glu Asp His Arg Gly Lys Thr Val Tyr Asp Val Ala Ser			
20	25	30	
Gly Asp Ala Leu Phe Ile Ser Glu Leu Gly Pro Leu Pro Glu Asn Val			
35	40	45	
Thr Trp Leu Ser Pro Glu Gly Glu Phe Gln Lys Trp Asn Gly Thr Ala			
50	55	60	
Trp Val Lys Asp Thr Glu Ala Glu Lys Leu Phe Arg Ile Arg Glu Ala			
65	70	75	80
Glu Glu Thr Lys Asn Ser Leu Met Gln Val Ala Ser Glu His Ile Ala			
85	90	95	
Pro Leu Gln Asp Ala Val Asp Leu Glu Ile Ala Thr Glu Glu Glu Thr			
100	105	110	
Ser Leu Leu Glu Ala Trp Lys Lys Tyr Arg Val Leu Leu Asn Arg Val			
115	120	125	
Asp Thr Ser Thr Ala Pro Asp Ile Glu Trp Pro Thr Asn Pro Val Arg			
130	135	140	
Glu			

-continued

145

```

<210> SEQ ID NO 5
<211> LENGTH: 3791
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic yej0 wildtype gene

<400> SEQUENCE: 5
atgcatcaat ctggttctgt ttctcttgt cgttccgcaa tatctgttct ggtggetaca    60
gcgttatgga aggtgcgaac aagtcctga tatgagatca tgtttgcac ctggagccat    120
agaacagggt tcatcatgag tcatcaactt accttcgccc acagtgaatt cagcagtaag    180
cgccgtcaga ccagaaaaga gattttcttg tcccgcattg agcagattct gccatggcaa    240
aacatggtgg aagtcacoga gccggtttac cccaaggctg gtaatggccc gcgaccttat    300
ccgctggaaa ccattgctacg cattcactgc atgcagcatt ggtacaaact gagcgatggc    360
gcgatggaag atgctctgta cgaaatcgcc tccatgctgc tgtttgcacc gttatcctg    420
gatagcgctc tgccggaccg caccaccatc atgaatttcc gccacctgct ggagcagcat    480
caactggccc gccaatgttt caagaccatc aatcgctggc tggccgaagc aggcgctcatg    540
atgactcaag gcaccttggc cgatgccacc atcattgagg caccagctc gaccaagaac    600
aaagagcagc aacgcgatcc ggagatgcat cagaccaaga aaggcaatca gtggcacttt    660
ggcatgaagg ccacatttgg tgcgatgcc aagagtggcc tgaccacagc cctggtcacc    720
accgcgcca acgagcatga cctcaatcag ctgggtaatc tgctgcatgg agaggagcaa    780
ttgtctcag ccgatgcccg ctaccaaggg gcgccacagc gcgaggagct ggccgaggtg    840
gatgtggact ggctgatgac cgagcgcccc ggcaaggtaa gaaccttgaa acagcatcca    900
cgcaagaaca aaacggccat caacatcgaa tacatgaaag ccagcatccg ggccaggggtg    960
gagcaccatc ttgcacatc caagcgacag ttcggcttcg tgaaagccag atacaagggg    1020
ttgctgaaaa acgataacca actggcgatg ttattcacgc tggccaaact gtttcgggcg    1080
gaccaaatag tacgtcagtg ggagagatct cactaaaaac tggggataac gccttaaatg    1140
gcgaagaaac ggtctaaata ggctgattca aggcatttac gggagaaaaa atcggetcaa    1200
acatgaagaa atgaaatgac tgagttagcc gagaagaatt tccccgctta ttgcacctt    1260
ccttatattc acccatagca ttggcatcaa ctggtgagta tggtagagaca gttgatggtg    1320
ttgtcctgga aaaagatatt cagctggttt atgggaccgc caataatcag aaaaatcaatc    1380
ctggcgggaga acagcatata aaagaatttg gtgtaagtaa taatactgaa attaacggag    1440
ggtatcagta cattgaaatg aatggcgccc cagaatactc agtattaaat gacggttatc    1500
aaattgttca aatgggtggc gcggcaaac agactacgct caataatggt gtgctacagg    1560
tttatggcgc agcgaatgat accacgatta aaggcgggcg cttaatcggt gaaaaagatg    1620
ggggggccgt ctttgcgctc atcgaaaagg gaggactact ggagggttaa gaggggggat    1680
ttgcatattg ggtagatcag aaagcaggcg gtgctattaa aacaaccacg cgggcatagg    1740
aggtattcgg aacaaaccgt ctcggtcagt tcgatataca gaatggtatt gctaataata    1800
tgttgttggg aaacggcgga agtttgcgag ttgaagaaaa tgacttcgct tataatacca    1860
ctgtagatag tggcggctta ctggagggtta tggatggcgg gactgtaact ggcggtgata    1920

```

-continued

```

aaaaagcagg cggaaaatta attgtctcaa cgaatgcgct ggaagtgagt ggtccaaaca 1980
gtaaaggcca atttagtata aaagatggtg tgtcaaaaaa ttatgaactg gatgatggtt 2040
cggggctcat tgttatggag gacacgcagg ccattgatac tacccttgat aagcatgcca 2100
ctatgcaatc gctgggaaag gatactggta cgaaagtgca ggcaaatgcg gtatatgatc 2160
tcggtcgatc atatcagaat ggaagtatca cgtattcctc aaaagccatc tctgaaaata 2220
tggttatcaa caatggccgc gctaacgtct gggctggcac aatggttaac gtttcagtca 2280
gagggaatga tggcattctt gaggtcatga agccgcaaat aaattatgca cccgcaatgt 2340
tgggtggtaa ggtagtgggt tctgagggcg cttcttttag aacgcatggt gccgtggata 2400
ccagcaaagc ggacgtttcg ctgaaaaata gcgtatggac catcattgcc gatataccta 2460
cgacgaacca aacaccctc ctcaacttag ccaaccctgc gatgtctgac gcaaatgtga 2520
ttatgatgga tgagccagtg actcgttcat cagtgcggc aagtgcggaa aatttcatta 2580
cgttgaccac caataccctg tcgggaaacg gcaattttta tatgcgtacc gatatggcta 2640
atcatcagag cgatcagctc aacgtcaccg gtcaggcaac aggtgatctc aaaatattcg 2700
tgacggacac cggtgccagc ccggcagcag gagatagcct tacactggta acaacgggcg 2760
gcggtgatgc tgcatttacg ttgggcaatg ccggaggcgt tgttgatatc ggtacgtatg 2820
aatatacctt gctggataat ggcaaccata gctggagtct ggcaagaaat cgcgcgcaaa 2880
ttacccttc aaccactgat gtgctgaata tggcggccgc acaaccgctg gtatttgatg 2940
cagaactgga caccgtcgtg gagcgtcttg gtacgtaaa aggcgtagt tacgatacgg 3000
cgatgtggag ttcggcaatt aacaccgca acaacgtgac cactgatgcg ggagctgggt 3060
ttgagcaaac attgacgggc ctgacgctcg gtatcgatag ccgtttctcc cgtgaagaaa 3120
gcagtacaat tcgcggcttg atctttgggt actctcattc tgatattggt tttgatcgcg 3180
gcgcaaaagg taatatcgat agctatacc tgggggctta tgccggttgg gagcatcaga 3240
acggtgccta tgttgatggg gtggtgaaag ttgaccgttt tgccaacacc atccatggca 3300
agatgagtaa tggggcaaca gcgtttggcg attacaatag taacggcgcg ggtgctcatg 3360
ttgagagcgg gttccgttgg gttgacggat tgtggagtgt tagaccctat ctggcctta 3420
cggctttac cacagatggt caggactaca cgttatcaaa cggcatgccc gcggatgtgg 3480
gaaataccg gatattacgc gctgaagcgg gaacggcggg aagctatcac atggacctgc 3540
aaaacggtac gacgctggaa ccctggctga aagcggcctg gcgtcaggaa tacgccgatt 3600
ctaaccaggt gaaagttaat gacgatggca aatttaataa tgatgtggct ggaaccagtg 3660
gcgtttatca ggtggtata aggtcatcgt ttaccocgac gttaagcggg catttgctag 3720
tcagctatgg caatggcgca ggggtagaat cgccgtggaa tactcaggcg ggtgtggtct 3780
ggacgttctg a 3791

```

```

<210> SEQ ID NO 6
<211> LENGTH: 1221
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic yeJ0 wildtype protein

```

```

<400> SEQUENCE: 6

```

```

Met His Gln Ser Gly Ser Val Ser Leu Cys Arg Ser Ala Ile Ser Val
1           5           10           15

```

-continued

Leu Val Ala Thr Ala Leu Trp Lys Val Arg Thr Ser Pro Tyr Glu Ile
20 25 30

Met Phe Val Ile Trp Ser His Arg Thr Gly Phe Ile Met Ser His Gln
35 40 45

Leu Thr Phe Ala Asp Ser Glu Phe Ser Ser Lys Arg Arg Gln Thr Arg
50 55 60

Lys Glu Ile Phe Leu Ser Arg Met Glu Gln Ile Leu Pro Trp Gln Asn
65 70 75 80

Met Val Glu Val Ile Glu Pro Phe Tyr Pro Lys Ala Gly Asn Gly Arg
85 90 95

Arg Pro Tyr Pro Leu Glu Thr Met Leu Arg Ile His Cys Met Gln His
100 105 110

Trp Tyr Asn Leu Ser Asp Gly Ala Met Glu Asp Ala Leu Tyr Glu Ile
115 120 125

Ala Ser Met Arg Leu Phe Ala Arg Leu Ser Leu Asp Ser Ala Leu Pro
130 135 140

Asp Arg Thr Thr Ile Met Asn Phe Arg His Leu Leu Glu Gln His Gln
145 150 155 160

Leu Ala Arg Gln Leu Phe Lys Thr Ile Asn Arg Trp Leu Ala Glu Ala
165 170 175

Gly Val Met Met Thr Gln Gly Thr Leu Val Asp Ala Thr Ile Ile Glu
180 185 190

Ala Pro Ser Ser Thr Lys Asn Lys Glu Gln Gln Arg Asp Pro Glu Met
195 200 205

His Gln Thr Lys Lys Gly Asn Gln Trp His Phe Gly Met Lys Ala His
210 215 220

Ile Gly Val Asp Ala Lys Ser Gly Leu Thr His Ser Leu Val Thr Thr
225 230 235 240

Ala Ala Asn Glu His Asp Leu Asn Gln Leu Gly Asn Leu Leu His Gly
245 250 255

Glu Glu Gln Phe Val Ser Ala Asp Ala Gly Tyr Gln Gly Ala Pro Gln
260 265 270

Arg Glu Glu Leu Ala Glu Val Asp Val Asp Trp Leu Ile Ala Glu Arg
275 280 285

Pro Gly Lys Val Arg Thr Leu Lys Gln His Pro Arg Lys Asn Lys Thr
290 295 300

Ala Ile Asn Ile Glu Tyr Met Lys Ala Ser Ile Arg Ala Arg Val Glu
305 310 315 320

His Pro Phe Arg Ile Ile Lys Arg Gln Phe Gly Phe Val Lys Ala Arg
325 330 335

Tyr Lys Gly Leu Leu Lys Asn Asp Asn Gln Leu Ala Met Leu Phe Thr
340 345 350

Leu Ala Asn Leu Phe Arg Ala Asp Gln Met Ile Arg Gln Trp Glu Arg
355 360 365

Ser His Lys Leu Gly Ile Thr Pro Met Ala Lys Lys Arg Ser Lys Ala
370 375 380

Asp Ser Arg His Leu Arg Glu Lys Lys Ser Ala Gln Thr Arg Asn Glu
385 390 395 400

Met Thr Glu Ser Ala Glu Lys Asn Phe Pro Ala Tyr Ser His Leu Pro
405 410 415

-continued

Tyr	Ile	His	Pro	His	Trp	His	Gln	Leu	Leu	Ser	Met	Val	Arg	Gln	Leu
			420					425					430		
Met	Val	Leu	Ser	Trp	Lys	Lys	Ile	Ser	Ser	Trp	Phe	Met	Gly	Pro	Pro
		435					440					445			
Ile	Ile	Arg	Lys	Ser	Ile	Leu	Ala	Glu	Asn	Ser	Ile	Lys	Asn	Leu	Val
450					455						460				
Val	Ile	Ile	Leu	Lys	Leu	Thr	Glu	Gly	Ile	Ser	Thr	Leu	Lys	Met	Ala
465				470						475					480
Pro	Gln	Asn	Thr	Gln	Tyr	Met	Thr	Val	Ile	Lys	Leu	Phe	Lys	Trp	Val
			485						490					495	
Ala	Arg	Gln	Thr	Arg	Leu	Arg	Ser	Ile	Met	Val	Cys	Tyr	Arg	Phe	Met
			500					505					510		
Ala	Gln	Arg	Met	Ile	Pro	Arg	Leu	Lys	Ala	Gly	Ala	Ser	Leu	Lys	Lys
		515					520					525			
Met	Gly	Gly	Pro	Ser	Leu	Ser	Leu	Ser	Lys	Arg	Glu	Asp	Tyr	Trp	Arg
530					535						540				
Leu	Lys	Arg	Gly	Asp	Leu	His	Leu	Arg	Ile	Arg	Lys	Gln	Ala	Val	Leu
545				550						555					560
Leu	Lys	Gln	Pro	Arg	Gly	Pro	Trp	Arg	Tyr	Ser	Glu	Gln	Thr	Val	Ser
			565						570					575	
Val	Ser	Ser	Ile	Ser	Arg	Met	Val	Leu	Leu	Ile	Ile	Cys	Cys	Trp	Lys
			580					585					590		
Thr	Ala	Glu	Val	Cys	Glu	Leu	Lys	Lys	Met	Thr	Ser	Leu	Ile	Ile	Pro
		595					600						605		
Leu	Ile	Val	Ala	Ala	Tyr	Trp	Arg	Leu	Trp	Met	Ala	Gly	Leu	Leu	Ala
610					615						620				
Leu	Ile	Lys	Lys	Gln	Ala	Glu	Asn	Leu	Ser	Gln	Arg	Met	Arg	Trp	Lys
625				630						635					640
Val	Val	Gln	Thr	Val	Lys	Ala	Asn	Leu	Val	Lys	Met	Val	Cys	Gln	Lys
			645						650					655	
Ile	Met	Asn	Trp	Met	Met	Val	Pro	Gly	Ser	Leu	Leu	Trp	Arg	Thr	Arg
		660						665					670		
Arg	Pro	Leu	Ile	Leu	Ser	Leu	Ile	Ser	Met	Pro	Leu	Cys	Asn	Arg	Trp
		675					680						685		
Glu	Arg	Ile	Leu	Val	Arg	Lys	Cys	Arg	Gln	Met	Arg	Tyr	Met	Ile	Ser
690					695						700				
Val	Asp	His	Ile	Arg	Met	Glu	Val	Ser	Arg	Ile	Pro	Gln	Lys	Pro	Ser
705				710						715					720
Leu	Lys	Ile	Trp	Leu	Ser	Thr	Met	Ala	Ala	Leu	Thr	Ser	Gly	Leu	Ala
			725						730					735	
Gln	Trp	Leu	Thr	Phe	Gln	Ser	Glu	Gly	Met	Met	Ala	Phe	Leu	Arg	Ser
			740					745					750		
Ser	Arg	Lys	Ile	Met	His	Pro	Gln	Cys	Trp	Trp	Val	Arg	Trp	Phe	Leu
		755					760						765		
Arg	Ala	Leu	Leu	Leu	Glu	Arg	Met	Val	Pro	Trp	Ile	Pro	Ala	Lys	Arg
		770				775						780			
Thr	Phe	Arg	Ser	Lys	Ile	Ala	Tyr	Gly	Pro	Ser	Leu	Pro	Ile	Ser	Leu
785				790						795					800
Arg	Arg	Thr	Lys	Thr	Pro	Ser	Ser	Thr	Pro	Thr	Leu	Arg	Cys	Leu	Thr
			805						810					815	
Gln	Met	Leu	Trp	Met	Ser	Gln	Leu	Val	His	Gln	Arg	Gln	Val	Arg	Lys

-continued

820					825					830					
Ile	Ser	Leu	Arg	Pro	Pro	Ile	Pro	Cys	Arg	Glu	Thr	Ala	Ile	Phe	Ile
		835					840					845			
Cys	Val	Pro	Ile	Trp	Leu	Ile	Ile	Arg	Ala	Ile	Ser	Ser	Thr	Ser	Pro
		850					855					860			
Val	Arg	Gln	Gln	Val	Ile	Ser	Lys	Tyr	Ser	Arg	Thr	Pro	Val	Pro	Ala
		865					870					875			880
Arg	Gln	Gln	Glu	Ile	Ala	Leu	His	Trp	Gln	Arg	Ala	Ala	Val	Met	Leu
				885					890					895	
His	Leu	Arg	Trp	Ala	Met	Pro	Glu	Ala	Leu	Leu	Ile	Ser	Val	Arg	Met
				900					905					910	
Asn	Ile	Pro	Cys	Trp	Ile	Met	Ala	Thr	Ile	Ala	Gly	Val	Trp	Gln	Arg
				915					920					925	
Ile	Ala	Arg	Lys	Leu	Pro	Leu	Gln	Pro	Leu	Met	Cys	Ile	Trp	Arg	Pro
				930					935					940	
His	Asn	Arg	Trp	Tyr	Leu	Met	Gln	Asn	Trp	Thr	Pro	Cys	Val	Ser	Val
				945					950					955	960
Leu	Val	Ala	Lys	Ala	Leu	Val	Thr	Ile	Arg	Arg	Cys	Gly	Val	Arg	Gln
				965					970					975	
Leu	Thr	Pro	Ala	Thr	Thr	Pro	Leu	Met	Arg	Glu	Leu	Val	Leu	Ser	Lys
				980					985					990	
His	Arg	Ala	Arg	Ser	Val	Ser	Ile	Ala	Val	Ser	Pro	Val	Lys	Lys	Ala
				995					1000					1005	
Val	Gln	Phe	Ala	Ala	Ser	Leu	Val	Thr	Leu	Ile	Leu	Ile	Leu	Val	
				1010					1015					1020	
Leu	Ile	Ala	Ala	Ala	Lys	Val	Ile	Ser	Ile	Ala	Ile	Pro	Trp	Gly	
				1025					1030					1035	
Leu	Met	Pro	Val	Gly	Ser	Ile	Arg	Thr	Val	Pro	Met	Leu	Met	Gly	
				1040					1045					1050	
Trp	Lys	Leu	Thr	Val	Leu	Pro	Thr	Pro	Ser	Met	Ala	Arg	Val	Met	
				1055					1060					1065	
Gly	Gln	Gln	Arg	Leu	Ala	Ile	Thr	Ile	Val	Thr	Ala	Arg	Val	Leu	
				1070					1075					1080	
Met	Leu	Arg	Ala	Gly	Ser	Val	Gly	Leu	Thr	Asp	Cys	Gly	Val	Leu	
				1085					1090					1095	
Asp	Pro	Ile	Trp	Pro	Leu	Pro	Ala	Leu	Pro	Gln	Met	Val	Arg	Thr	
				1100					1105					1110	
Thr	Arg	Tyr	Gln	Thr	Ala	Cys	Ala	Arg	Met	Trp	Glu	Ile	Pro	Gly	
				1115					1120					1125	
Tyr	Tyr	Ala	Leu	Lys	Arg	Glu	Arg	Arg	Ala	Ile	Thr	Trp	Thr	Cys	
				1130					1135					1140	
Lys	Thr	Val	Arg	Arg	Trp	Asn	Pro	Gly	Lys	Arg	Pro	Cys	Val	Arg	
				1145					1150					1155	
Asn	Thr	Pro	Ile	Leu	Thr	Arg	Lys	Leu	Met	Thr	Met	Ala	Asn	Leu	
				1160					1165					1170	
Ile	Met	Met	Trp	Leu	Glu	Pro	Val	Ala	Phe	Ile	Arg	Leu	Val	Gly	
				1175					1180					1185	
His	Arg	Leu	Pro	Arg	Arg	Ala	Val	Ile	Cys	Gln	Ser	Ala	Met	Ala	
				1190					1195					1200	
Met	Ala	Gln	Gly	Asn	Arg	Arg	Gly	Ile	Leu	Arg	Arg	Val	Trp	Ser	
				1205					1210					1215	

-continued

Gly Arg Ser
1220

<210> SEQ ID NO 7
<211> LENGTH: 3791
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic yej0 mutated gene

<400> SEQUENCE: 7

```

atgcatcaat ctggttctgt ttctctttgt cgttccgcaa tatctgttct ggtggctaca      60
gcgtaaagga aggtgcgaac aagtcctga tatgagatca tgtttgcac ctggagccat      120
agaacagggt tcatcatgag tcatcaactt accttcgccc acagtgaatt cagcagtaag      180
cgccgtcaga ccagaaaaga gattttcttg tcccgcattg agcagattct gccatggcaa      240
aacatggtgg aagtcacoga gccgttttac cccaaggctg gtaatggccg gcgaccttat      300
ccgctggaaa ccatgctacg cattcaactg atgcagcatt ggtacaacct gagcgatggc      360
gcgatggaag atgctctgta cgaaatcgcc tccatgcgct tgtttgcccg gttatccctg      420
gatagcgccct tgccggaccg caccaccatc atgaatttcc gccacctgct ggagcagcat      480
caactggccc gccaatggtt caagaccatc aatcgctggc tggccgaagc aggcgcatg      540
atgactcaag gcaccttggc cgatgccacc atcattgagg caccagctc gaccaagaac      600
aaagagcagc aaccgcatcc ggagatgcat cagaccaaga aaggcaatca gtggcacttt      660
ggcatgaagg cccacattgg tgcgatgcc aagagtggcc tgaccacag cctggtcacc      720
accgcgcca acgagcatga cctcaatcag ctgggtaatc tgctgcatgg agaggagcaa      780
tttgtctcag ccgatgcccg ctaccaaggg gcgccacagc gcgaggagct ggccgaggtg      840
gatgtggact ggctgatgct cgagcgcgcc ggcaaggtaa gaacctgaa acagcatcca      900
cgcaagaaca aaacggccat caacatcgaa tacatgaaag ccagcatccc ggccagggtg      960
gagcaccatc ttcgcatcat caagcgacag ttcggcttcg tgaaagccag atacaagggg      1020
ttgctgaaaa acgataacca actggcgatg ttattcacgc tggccaacct gtttcgggcg      1080
gaccaaataa tacgtcagtg ggagagatct cactaaaaac tggggataac gccttaaatg      1140
gcaagaatac ggtctaaata ggctgattca aggcatttac gggagaaaaa atcggctcaa      1200
acatgaagaa atgaaatgac tgagtcagcc gagaagaatt tccccgctta ttcgacctt      1260
ccttatattc acccatagca ttggcatcaa ctggtgagta tgggtgagaca gttgatggtg      1320
ttgtcctgga aaaagatata cagctgggtt atgggaccgc caataatacg aaaaatcaatc      1380
ctggcggaga acagcatata aaagaatttg gtgtaagtaa taatactgaa attaacggag      1440
ggtatcagta cattgaaatg aatggcgccg cagaatactc agtattaaat gacggttatc      1500
aaattgttca aatgggtggc gcgcaaac accactacgct caataatggt gtgctacagg      1560
tttatggcgc agcgaatgat accacgatta aaggcgggcy cttaatcgtt gaaaaagatg      1620
ggggggccgt ctttctcctc atcgaaaagg gaggactact ggagggtaaa gaggggggat      1680
ttgcatctgc ggtagatcag aaagcaggcg gtgctattaa aacaaccacg cgggccatgg      1740
aggtattcgg aacaaaccgt ctcggtcagc tcgatataca gaatggtatt gctaataata      1800
tggtgttggg aaacggcgga agtttgcgag ttgaagaaaa tgacttcgct tataatacca      1860

```

-continued

```

ctgtagatag tggcggctta ctggaggtta tggatggcgg gactgtaact ggcggtgata 1920
aaaaagcagg cggaaaatta attgtctcaa cgaatgcgct ggaagtgagt ggtccaaaca 1980
gtaaaggcca atttagtata aaagatggtg tgtcaaaaa ttatgaactg gatgatggtt 2040
ccgggctcat tgttatggag gacacgcagg ccattgatac tatccttgat aagcatgcca 2100
ctatgcaatc gctgggaaag gatactggta cgaaagtgca ggcaaatgcg gtatatgatc 2160
tcggtcgcgc atatcagaat ggaagtatca cgtattcctc aaaagccatc tctgaaaata 2220
tggttatcaa caatggcgcg gctaacgtct gggctggcac aatggttaac gtttcagtca 2280
gagggaaatga tggcattcct gaggtcatga agccgcaaat aaattatgca cccgcaatgt 2340
tgggtggtaa ggtagtgggt tctgagggcg cttcttttag aacgcatggt gccgtggata 2400
ccagcaaagc ggacgtttcg ctcgaaaata gcgatggac catcattgcc gatataccta 2460
cgacgaacca aaacacctc ctcaacttag ccaaccttgc gatgtctgac gcaaatgta 2520
ttatgatgga tgagccagt actcgttcat cagtgcggc aagtgcggaa aatttcatta 2580
cgttgaccac caataccctg tccggaaacg gcaattttta tatgcgtacc gatatggcta 2640
atcatcagag cgatcagctc aacgtcaccg gtcaggcaac aggtgatttc aaaatattcg 2700
tgacggacac cgggtgccgc cgggcagcag gagatagcct tacactggta acaacgggcg 2760
gcggtgatgc tgcatttacg ttgggcaatg ccggaggcgt tgttgatata ggtacgatg 2820
aatatacctt gctggataat ggcaaccata gctggagtct ggcaagaaat cgcgcgcaaa 2880
ttacccttc aaccactgat gtgctgaata tggcggcgc acaaccgctg gtatttgatg 2940
cagaactgga caccgtcgtg gagcgtcttg gttagcgtaaa aggcgttagt tacgatacgg 3000
cgatgtggag ttcggcaatt aacaccgca acaacgtgac cactgatgcg ggagctggtt 3060
ttgagcaaac attgacgggc ctgacgctcg gtatcgatag ccgtttctcc cgtgaagaaa 3120
gcagtacaat tccggccttg atctttggtt actctcattc tgatattggt tttgatcgcg 3180
gcggcaaaag taatatcgat agctataccc tgggggctta tgccggttgg gagcatcaga 3240
acggtgccta tgttgatggg gtggtgaaag ttgaccggtt tgccaacacc atccatggca 3300
agatgagtaa tggggcaaca gcgtttggcg attacaatag taacggcgcg ggtgctcatg 3360
ttgagagcgg gttccgctgg gttgacggat tgtggagtgt tagaccetat ctggccttta 3420
ccggctttac cacagatggt caggactaca cgttatcaaa cggcatgccc gcggatgtgg 3480
gaaataccgc gatattacgc gctgaagcgg gaacggcggg aagctatcac atggacctgc 3540
aaaacggtac gacgctggaa cctggctga aagcggcggg gcgtcaggaa tacgcccatt 3600
ctaaccaggg gaaagttaat gacgatggca aatttaataa tgatgtggct ggaaccagtg 3660
gcgtttatca ggctggata aggtcatcgt ttaccccgac gtaagcggg catttgcag 3720
tcagctatgg caatggcgca ggggtagaat cgccgtggaa tactcaggcg ggtgtggtct 3780
ggacgttctg a 3791

```

<210> SEQ ID NO 8

<211> LENGTH: 1221

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic yeJ0 mutated protein

<400> SEQUENCE: 8

-continued

Met	His	Gln	Ser	Gly	Ser	Val	Ser	Leu	Cys	Arg	Ser	Ala	Ile	Ser	Val	1	5	10	15
Leu	Val	Ala	Thr	Ala	Leu	Arg	Lys	Val	Arg	Thr	Ser	Pro	Tyr	Glu	Ile	20	25	30	
Met	Phe	Val	Ile	Trp	Ser	His	Arg	Thr	Gly	Phe	Ile	Met	Ser	His	Gln	35	40	45	
Leu	Thr	Phe	Ala	Asp	Ser	Glu	Phe	Ser	Ser	Lys	Arg	Arg	Gln	Thr	Arg	50	55	60	
Lys	Glu	Ile	Phe	Leu	Ser	Arg	Met	Glu	Gln	Ile	Leu	Pro	Trp	Gln	Asn	65	70	75	80
Met	Val	Glu	Val	Ile	Glu	Pro	Phe	Tyr	Pro	Lys	Ala	Gly	Asn	Gly	Arg	85	90	95	
Arg	Pro	Tyr	Pro	Leu	Glu	Thr	Met	Leu	Arg	Ile	His	Cys	Met	Gln	His	100	105	110	
Trp	Tyr	Asn	Leu	Ser	Asp	Gly	Ala	Met	Glu	Asp	Ala	Leu	Tyr	Glu	Ile	115	120	125	
Ala	Ser	Met	Arg	Leu	Phe	Ala	Arg	Leu	Ser	Leu	Asp	Ser	Ala	Leu	Pro	130	135	140	
Asp	Arg	Thr	Thr	Ile	Met	Asn	Phe	Arg	His	Leu	Leu	Glu	Gln	His	Gln	145	150	155	160
Leu	Ala	Arg	Gln	Leu	Phe	Lys	Thr	Ile	Asn	Arg	Trp	Leu	Ala	Glu	Ala	165	170	175	
Gly	Val	Met	Met	Thr	Gln	Gly	Thr	Leu	Val	Asp	Ala	Thr	Ile	Ile	Glu	180	185	190	
Ala	Pro	Ser	Ser	Thr	Lys	Asn	Lys	Glu	Gln	Gln	Arg	Asp	Pro	Glu	Met	195	200	205	
His	Gln	Thr	Lys	Lys	Gly	Asn	Gln	Trp	His	Phe	Gly	Met	Lys	Ala	His	210	215	220	
Ile	Gly	Val	Asp	Ala	Lys	Ser	Gly	Leu	Thr	His	Ser	Leu	Val	Thr	Thr	225	230	235	240
Ala	Ala	Asn	Glu	His	Asp	Leu	Asn	Gln	Leu	Gly	Asn	Leu	Leu	His	Gly	245	250	255	
Glu	Glu	Gln	Phe	Val	Ser	Ala	Asp	Ala	Gly	Tyr	Gln	Gly	Ala	Pro	Gln	260	265	270	
Arg	Glu	Glu	Leu	Ala	Glu	Val	Asp	Val	Asp	Trp	Leu	Ile	Ala	Glu	Arg	275	280	285	
Pro	Gly	Lys	Val	Arg	Thr	Leu	Lys	Gln	His	Pro	Arg	Lys	Asn	Lys	Thr	290	295	300	
Ala	Ile	Asn	Ile	Glu	Tyr	Met	Lys	Ala	Ser	Ile	Arg	Ala	Arg	Val	Glu	305	310	315	320
His	Pro	Phe	Arg	Ile	Ile	Lys	Arg	Gln	Phe	Gly	Phe	Val	Lys	Ala	Arg	325	330	335	
Tyr	Lys	Gly	Leu	Leu	Lys	Asn	Asp	Asn	Gln	Leu	Ala	Met	Leu	Phe	Thr	340	345	350	
Leu	Ala	Asn	Leu	Phe	Arg	Ala	Asp	Gln	Met	Ile	Arg	Gln	Trp	Glu	Arg	355	360	365	
Ser	His	Lys	Leu	Gly	Ile	Thr	Pro	Met	Ala	Lys	Lys	Arg	Ser	Lys	Ala	370	375	380	
Asp	Ser	Arg	His	Leu	Arg	Glu	Lys	Lys	Ser	Ala	Gln	Thr	Arg	Asn	Glu	385	390	395	400
Met	Thr	Glu	Ser	Ala	Glu	Lys	Asn	Phe	Pro	Ala	Tyr	Ser	His	Leu	Pro				

-continued

			405					410				415			
Tyr	Ile	His	Pro	His	Trp	His	Gln	Leu	Leu	Ser	Met	Val	Arg	Gln	Leu
			420					425					430		
Met	Val	Leu	Ser	Trp	Lys	Lys	Ile	Ser	Ser	Trp	Phe	Met	Gly	Pro	Pro
		435					440					445			
Ile	Ile	Arg	Lys	Ser	Ile	Leu	Ala	Glu	Asn	Ser	Ile	Lys	Asn	Leu	Val
	450					455					460				
Val	Ile	Ile	Leu	Lys	Leu	Thr	Glu	Gly	Ile	Ser	Thr	Leu	Lys	Met	Ala
465					470					475					480
Pro	Gln	Asn	Thr	Gln	Tyr	Met	Thr	Val	Ile	Lys	Leu	Phe	Lys	Trp	Val
			485						490					495	
Ala	Arg	Gln	Thr	Arg	Leu	Arg	Ser	Ile	Met	Val	Cys	Tyr	Arg	Phe	Met
			500					505					510		
Ala	Gln	Arg	Met	Ile	Pro	Arg	Leu	Lys	Ala	Gly	Ala	Ser	Leu	Lys	Lys
		515					520						525		
Met	Gly	Gly	Pro	Ser	Leu	Ser	Leu	Ser	Lys	Arg	Glu	Asp	Tyr	Trp	Arg
	530					535					540				
Leu	Lys	Arg	Gly	Asp	Leu	His	Leu	Arg	Ile	Arg	Lys	Gln	Ala	Val	Leu
545					550					555					560
Leu	Lys	Gln	Pro	Arg	Gly	Pro	Trp	Arg	Tyr	Ser	Glu	Gln	Thr	Val	Ser
				565					570						575
Val	Ser	Ser	Ile	Ser	Arg	Met	Val	Leu	Leu	Ile	Ile	Cys	Cys	Trp	Lys
			580					585						590	
Thr	Ala	Glu	Val	Cys	Glu	Leu	Lys	Lys	Met	Thr	Ser	Leu	Ile	Ile	Pro
		595					600						605		
Leu	Ile	Val	Ala	Ala	Tyr	Trp	Arg	Leu	Trp	Met	Ala	Gly	Leu	Leu	Ala
	610					615					620				
Leu	Ile	Lys	Lys	Gln	Ala	Glu	Asn	Leu	Ser	Gln	Arg	Met	Arg	Trp	Lys
625				630						635					640
Val	Val	Gln	Thr	Val	Lys	Ala	Asn	Leu	Val	Lys	Met	Val	Cys	Gln	Lys
				645					650						655
Ile	Met	Asn	Trp	Met	Met	Val	Pro	Gly	Ser	Leu	Leu	Trp	Arg	Thr	Arg
			660					665						670	
Arg	Pro	Leu	Ile	Leu	Ser	Leu	Ile	Ser	Met	Pro	Leu	Cys	Asn	Arg	Trp
		675					680						685		
Glu	Arg	Ile	Leu	Val	Arg	Lys	Cys	Arg	Gln	Met	Arg	Tyr	Met	Ile	Ser
	690					695					700				
Val	Asp	His	Ile	Arg	Met	Glu	Val	Ser	Arg	Ile	Pro	Gln	Lys	Pro	Ser
705					710					715					720
Leu	Lys	Ile	Trp	Leu	Ser	Thr	Met	Ala	Ala	Leu	Thr	Ser	Gly	Leu	Ala
				725					730						735
Gln	Trp	Leu	Thr	Phe	Gln	Ser	Glu	Gly	Met	Met	Ala	Phe	Leu	Arg	Ser
			740					745					750		
Ser	Arg	Lys	Ile	Met	His	Pro	Gln	Cys	Trp	Trp	Val	Arg	Trp	Phe	Leu
		755					760						765		
Arg	Ala	Leu	Leu	Leu	Glu	Arg	Met	Val	Pro	Trp	Ile	Pro	Ala	Lys	Arg
	770					775							780		
Thr	Phe	Arg	Ser	Lys	Ile	Ala	Tyr	Gly	Pro	Ser	Leu	Pro	Ile	Ser	Leu
785					790					795					800
Arg	Arg	Thr	Lys	Thr	Pro	Ser	Ser	Thr	Pro	Thr	Leu	Arg	Cys	Leu	Thr
				805					810						815

-continued

Gln Met Leu Trp Met Ser Gln Leu Val His Gln Arg Gln Val Arg Lys
 820 825 830

Ile Ser Leu Arg Pro Pro Ile Pro Cys Arg Glu Thr Ala Ile Phe Ile
 835 840 845

Cys Val Pro Ile Trp Leu Ile Ile Arg Ala Ile Ser Ser Thr Ser Pro
 850 855 860

Val Arg Gln Gln Val Ile Ser Lys Tyr Ser Arg Thr Pro Val Pro Ala
 865 870 875 880

Arg Gln Gln Glu Ile Ala Leu His Trp Gln Arg Ala Ala Val Met Leu
 885 890 895

His Leu Arg Trp Ala Met Pro Glu Ala Leu Leu Ile Ser Val Arg Met
 900 905 910

Asn Ile Pro Cys Trp Ile Met Ala Thr Ile Ala Gly Val Trp Gln Arg
 915 920 925

Ile Ala Arg Lys Leu Pro Leu Gln Pro Leu Met Cys Ile Trp Arg Pro
 930 935 940

His Asn Arg Trp Tyr Leu Met Gln Asn Trp Thr Pro Cys Val Ser Val
 945 950 955 960

Leu Val Ala Lys Ala Leu Val Thr Ile Arg Arg Cys Gly Val Arg Gln
 965 970 975

Leu Thr Pro Ala Thr Thr Pro Leu Met Arg Glu Leu Val Leu Ser Lys
 980 985 990

His Arg Ala Arg Ser Val Ser Ile Ala Val Ser Pro Val Lys Lys Ala
 995 1000 1005

Val Gln Phe Ala Ala Ser Leu Val Thr Leu Ile Leu Ile Leu Val
 1010 1015 1020

Leu Ile Ala Ala Ala Lys Val Ile Ser Ile Ala Ile Pro Trp Gly
 1025 1030 1035

Leu Met Pro Val Gly Ser Ile Arg Thr Val Pro Met Leu Met Gly
 1040 1045 1050

Trp Lys Leu Thr Val Leu Pro Thr Pro Ser Met Ala Arg Val Met
 1055 1060 1065

Gly Gln Gln Arg Leu Ala Ile Thr Ile Val Thr Ala Arg Val Leu
 1070 1075 1080

Met Leu Arg Ala Gly Ser Val Gly Leu Thr Asp Cys Gly Val Leu
 1085 1090 1095

Asp Pro Ile Trp Pro Leu Pro Ala Leu Pro Gln Met Val Arg Thr
 1100 1105 1110

Thr Arg Tyr Gln Thr Ala Cys Ala Arg Met Trp Glu Ile Pro Gly
 1115 1120 1125

Tyr Tyr Ala Leu Lys Arg Glu Arg Arg Ala Ile Thr Trp Thr Cys
 1130 1135 1140

Lys Thr Val Arg Arg Trp Asn Pro Gly Lys Arg Pro Cys Val Arg
 1145 1150 1155

Asn Thr Pro Ile Leu Thr Arg Lys Leu Met Thr Met Ala Asn Leu
 1160 1165 1170

Ile Met Met Trp Leu Glu Pro Val Ala Phe Ile Arg Leu Val Gly
 1175 1180 1185

His Arg Leu Pro Arg Arg Ala Val Ile Cys Gln Ser Ala Met Ala
 1190 1195 1200

-continued

Met	Ala	Gln	Gly	Asn	Arg	Arg	Gly	Ile	Leu	Arg	Arg	Val	Trp	Ser
	1205					1210						1215		

Gly	Arg	Ser
	1220	

1. A method for generating microorganisms resistant to infection by phages wherein said method comprises:

- a) Growing the microorganism of interest until it reaches an optical density (600 nm) from 0.6 to 5;
- b) Adding a mixture of phages with a concentration greater than 100 plaque-forming units;
- c) Contacting the microorganism culture with phages from 30 minutes to 6 hours at room temperature;
- d) Allowing for cellular lysis from 1 hour to 8 hours at 37° C.;
- e) Allowing the culture to recover from 1 hour to 24 hours;
- f) Isolating colonies into plates with M9 medium; and
- g) Verifying the resistance of the microorganism by repeating steps from a) to f).

2. The method according to claim 1, wherein microorganisms resistant to infection by phages from the Siphoviridae and Myoviridae families are generated.

3. The method according to claim 2, wherein microorganisms are resistant to infection by phages λ , $\phi 80$ and T4 are generated.

4. The method according to claim 3, wherein the microorganism is an enterobacterium.

5. The method according to claim 4, wherein the enterobacterium is *Escherichia coli* strain LCT-BF-01.

6. A microorganism resistant to phage infection obtained by the method of claim 1.

7. A microorganism according to claim 6, wherein the microorganism is *Escherichia coli*

8. A microorganism according to claim 7, wherein the microorganism has mutations in *tfaD* and *yeyO* genes.

9. A microorganism according to claim 8, wherein the microorganism is resistant to the infection by phages from the Siphoviridae and Myoviridae families.

10. A microorganism according to claim 9, wherein the microorganism is resistant to the infection by phages λ , $\phi 80$ and T4.

11. The microorganism according to claim 10, wherein the microorganism is strain LCT-BF-01.

12. The microorganism according to claim 11, wherein said microorganism retains the same kinetic constants as a wild type strain.

* * * * *